



CRYSTAL DIGITAL PCR™ DETECTION KIT FOR SARS-COV-2

Romain Parillaud, PhD
Field Application Specialist
Stilla Technologies Inc.

Current molecular diagnostic tool to diagnose SARS-CoV-2

- Clinical symptoms difficult to dissociate from other respiratory infections
- Detection of SARS-CoV-2 Nucleic acid sequence

Real-Time PCR

- Patient having pneumonia and CT abnormalities but be initially RT-qPCR negative for SARS-CoV-2
=> Only later after onset observed positive on RT-PCR

Ai T, et al. Radiology, 2020.

Kanne JP, et al. Radiology, 2020

Yang W, et al. Radiology, 2020.

Winichakoon P, et.al. Journal of Clinical Microbiology Feb 2020

- RT-qPCR reported to be 50-70% sensitive

Arima Y, Emerg Infect Dis. 2020

Wu J, et al. Clin Infect Dis 2020

Fengting Yu, et al. Clin Infect Dis. 2020

Xie X, et.al. Radiology. 2020

Necessity to develop more sensitive detection tools

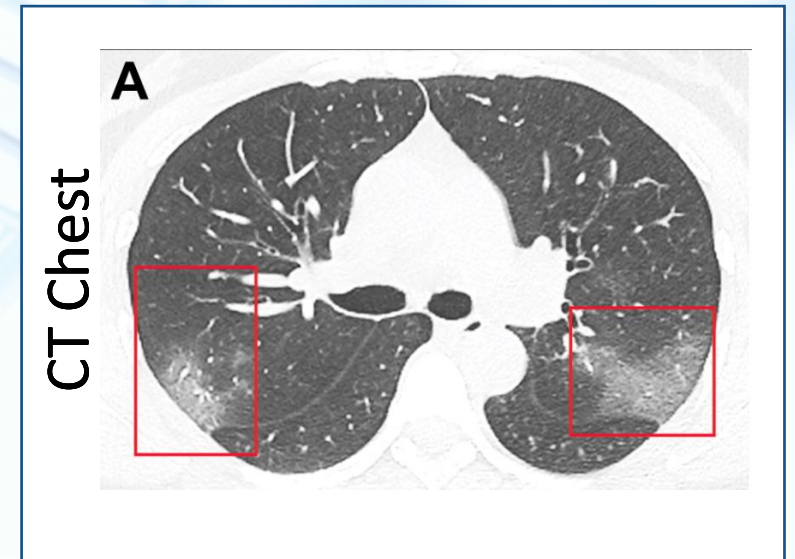


Image: <https://www.itnonline.com/article/ct-imaging-2019-novel-coronavirus-2019-ncov-pneumonia>

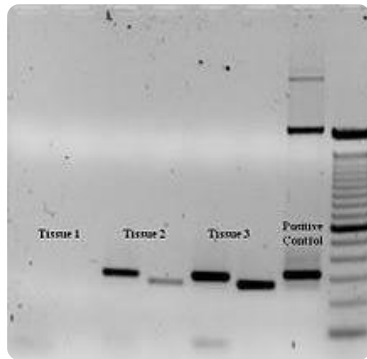


NEXT GENERATION OF PCR

Digital PCR

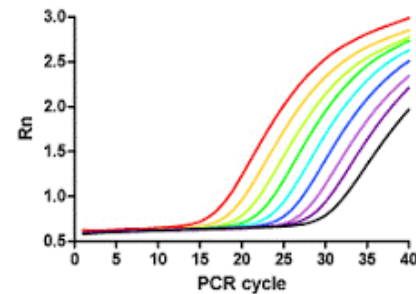
OUR MISSION:
MAKE DIGITAL PCR A LAB COMMODITY

PCR



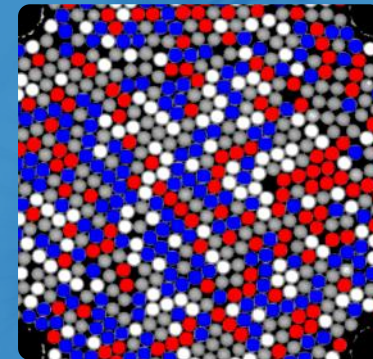
Amplify Target DNA

Quantitative PCR



Relative quantification
Real-time with standard curves
Ubiquitously spread method

Digital PCR



Absolute quantification
No standard curve
Increased sensitivity
Higher precision



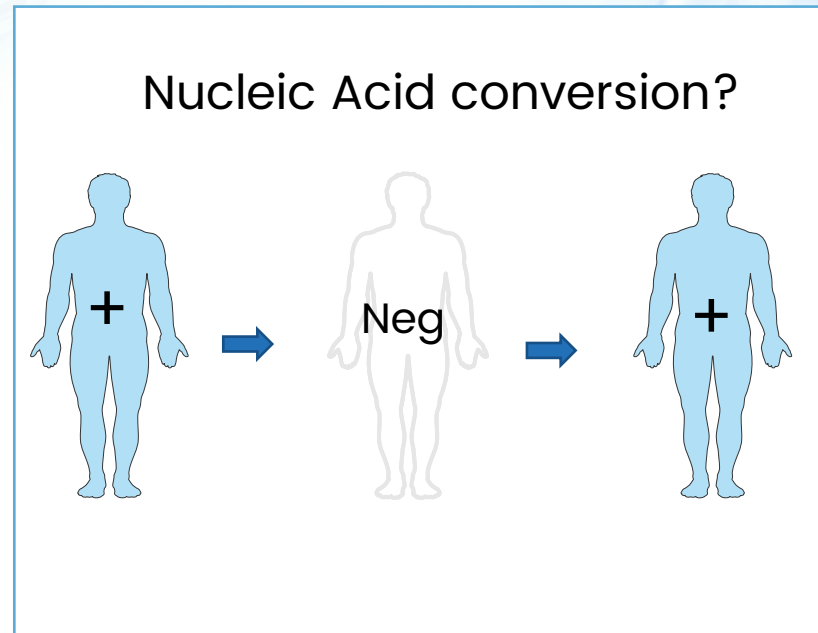
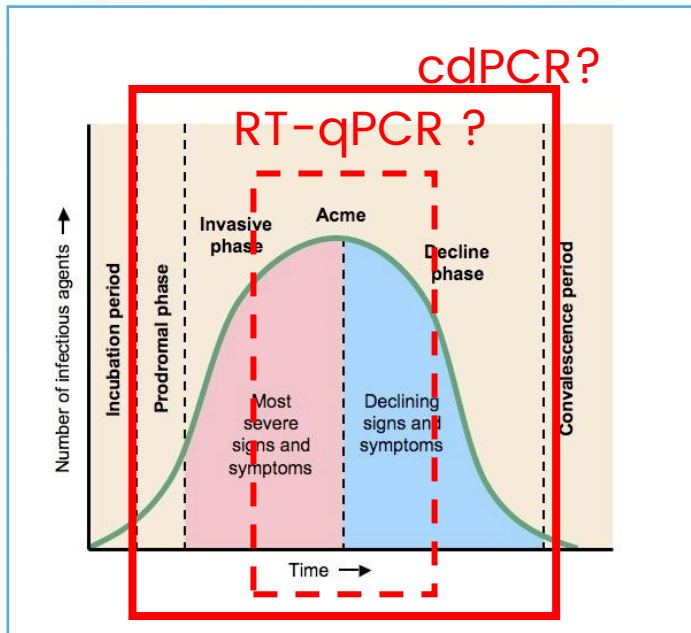
NEXT GENERATION OF PCR

Digital PCR

OUR MISSION:
MAKE DIGITAL PCR A LAB COMMODITY

- Digital PCR could be a valuable asset in the COVID-19 battle

Sensitivity



Ai Tang Xiao, Yi Xin Tong, Sheng Zhang, , J Med Virol. 2020 Apr 9

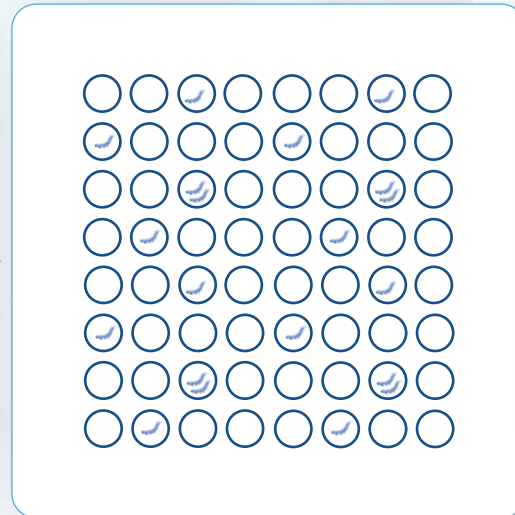
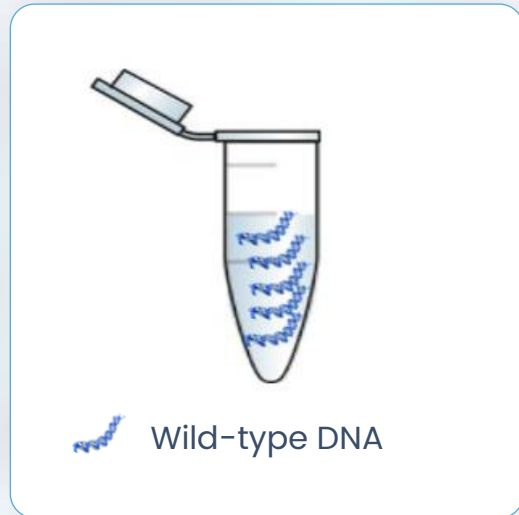
Digital PCR

Absolute quantification
No standard curve
Increased sensitivity
Higher precision

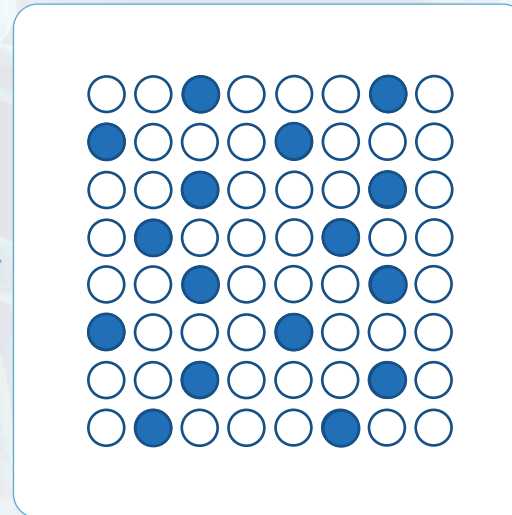


PRINCIPLE OF DIGITAL PCR

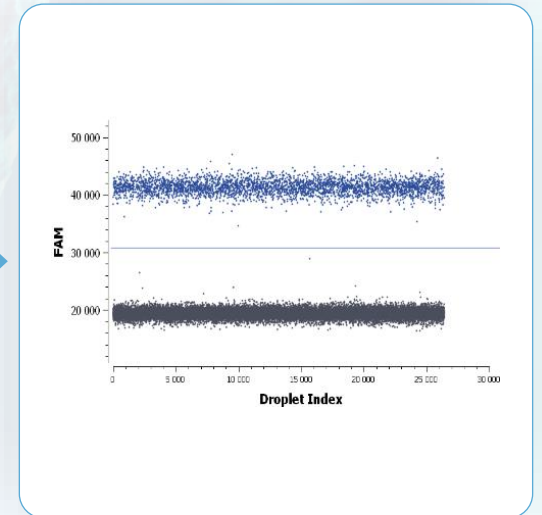
PARTITIONING



PCR



READING & ANALYSIS



RESULTS
2636 cp/μL with 2.2 %
uncertainty

POISSON STATISTICS

$$\frac{N_{pos}}{N_{tot}}$$



Naica™ System Workflow Crystal Digital PCR™

Sapphire Chip
(consumable)

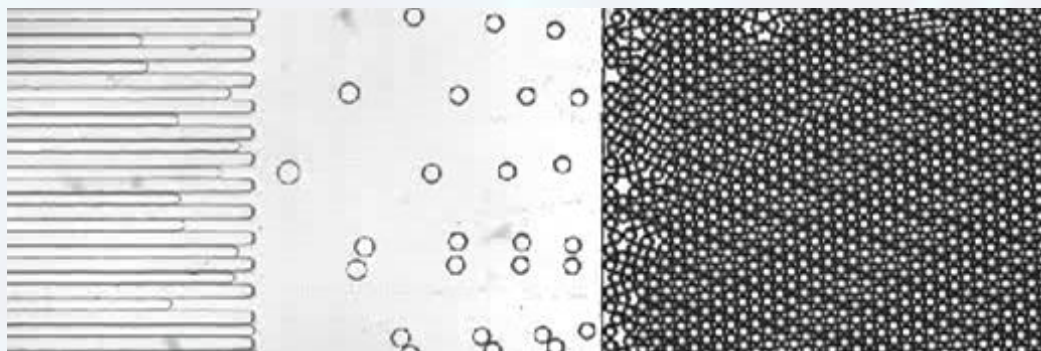


Naica™ Geode

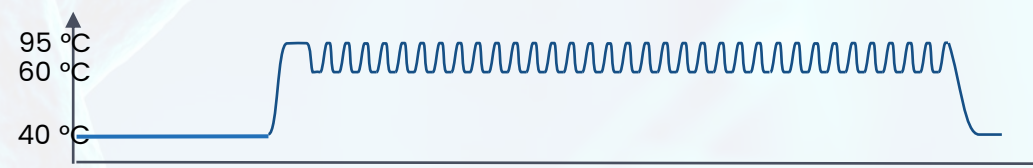
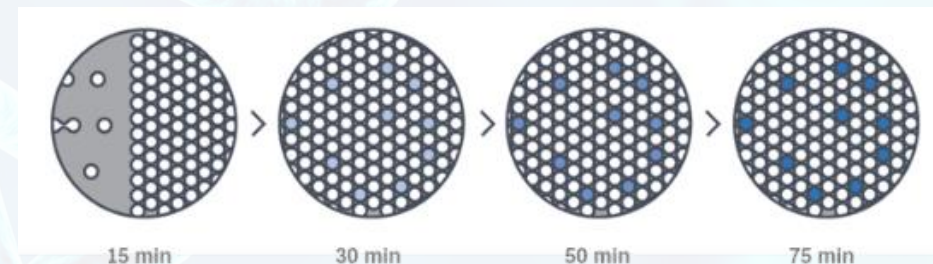


Step 2.1 – Partition

- 1-3 chips and 1-12 samples / run
- Contactless fluid injection

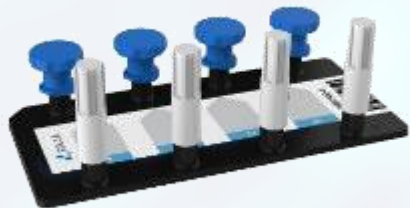


Step 2.2 – Amplify



Naica™ System Workflow Crystal Digital PCR™

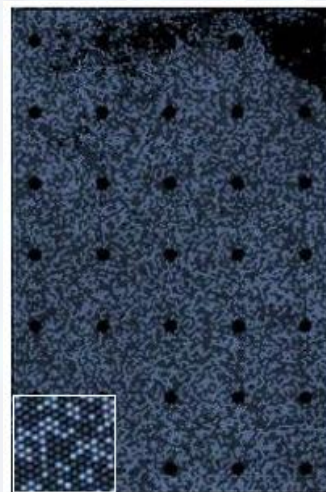
Sapphire Chip
(consumable)



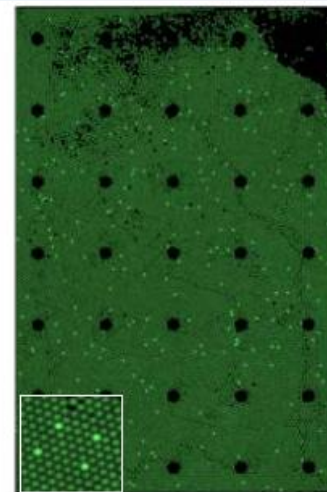
Naica™ Geode



Naica™ Prism3



Blue
Ex: 415-480 nm
Em: 495-520 nm
FAM...



Green
Ex: 530-550 nm
Em: 560-610 nm
HEX...



Red
Ex: 615-645 nm
Em: 655-720 nm
Cy@5...



Naica™ System Workflow Crystal Digital PCR™

Sapphire Chip
(consumable)



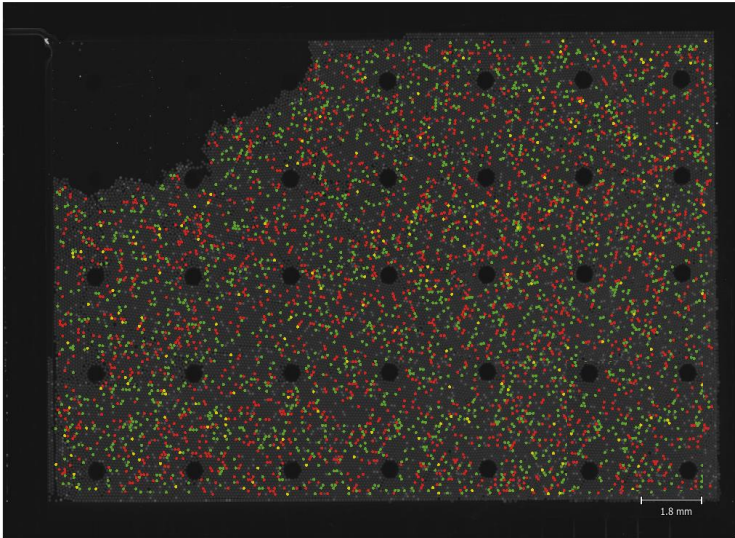
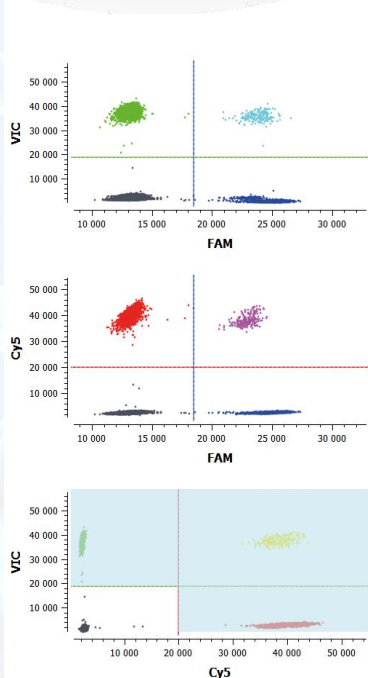
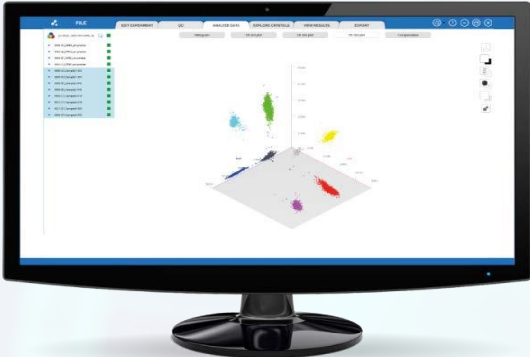
Naica™ Geode



Naica™ Prism3



Crystal Miner™ (software)



Color Mode: Auto Blue Cy5

Channel Selection: FAM Cy5 Cy3

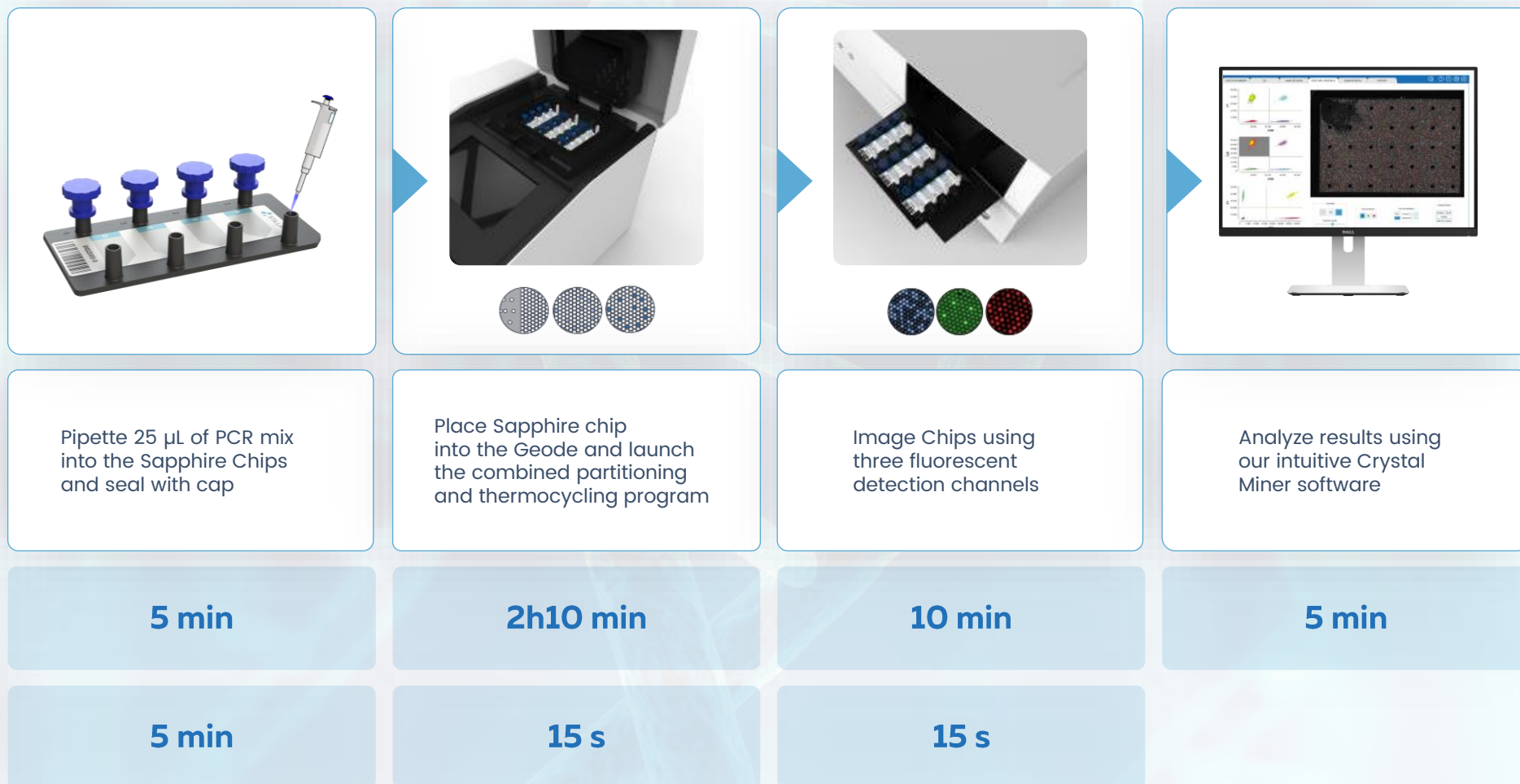
Population opacity:

Contrast Adjustment: Auto Contrast Reset Brightness

Droplet Exclusion:



PERFORM CRYSTAL DIGITAL PCR™ IN 2H30 WITH MINIMUM HANDS-ON TIME

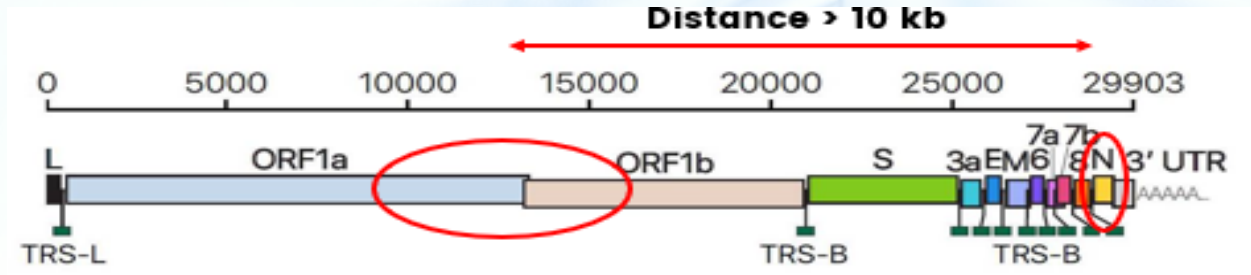


A novel kit for COVID-19 detection in human samples

Taking advantage of multiplexing for increase sensitivity

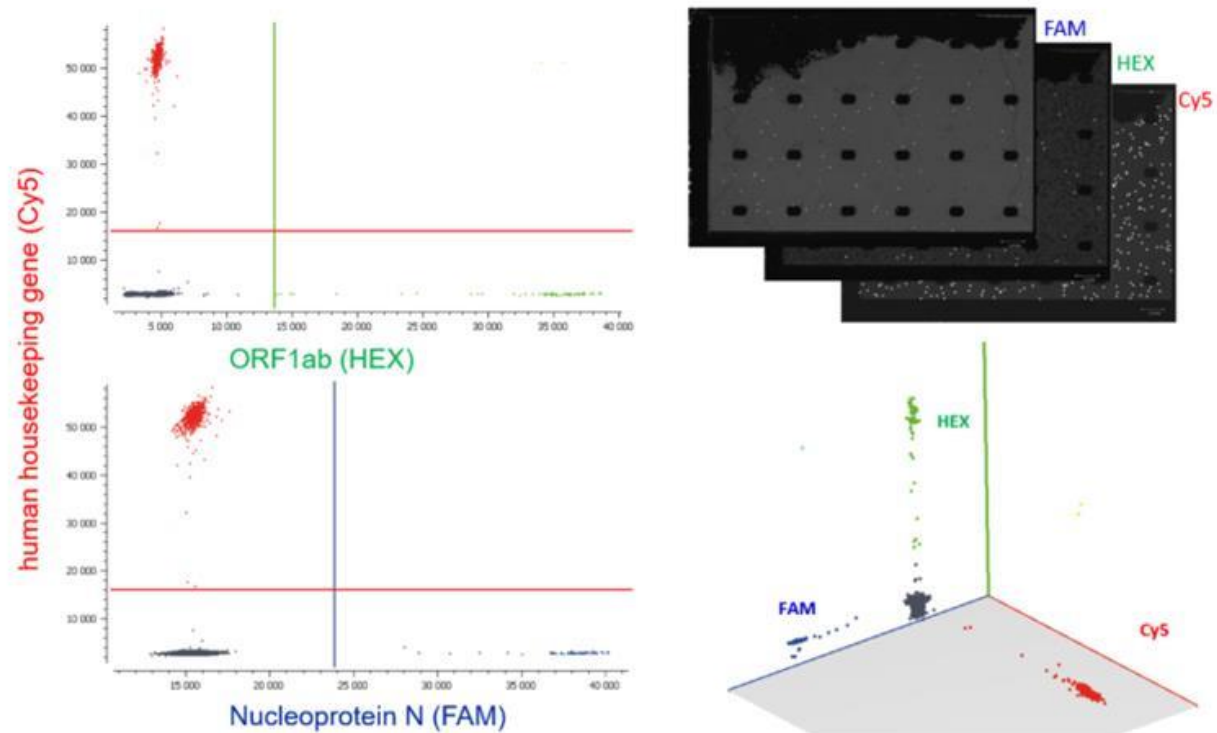
3-color kit to detect viral and human genes:

- COVID-19 ORF1ab (HEX)
- COVID-19 nucleoprotein N (FAM)
- Human control housekeeping gene (Cy5)



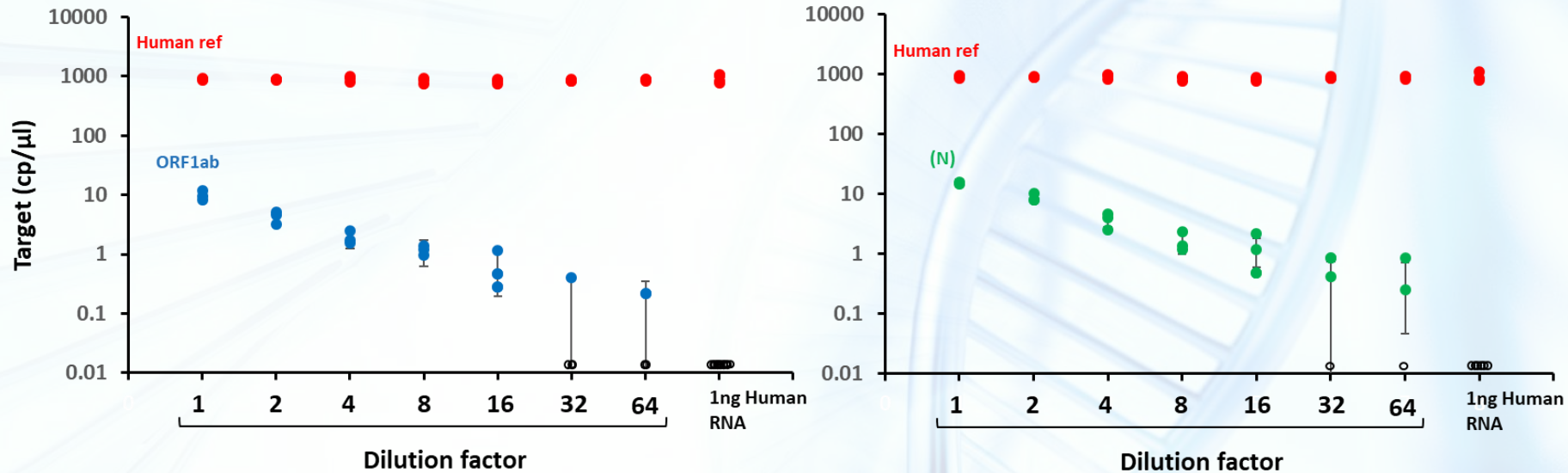
Kit components

- dPCR master mix1
- dPCR master mix2
- Primer and probe mix
- COVID-19 positive control
- COVID-19 negative control
- Sapphire Chips



Sensitive and specific detection of COVID-19 sequences

- A positive control containing ORF1ab and nucleoprotein N sequences was serially diluted and tested in triplicate.
- A total of 8 μl of positive control was assessed in a 25 μl reaction in a background of 1 ng of human RNA.

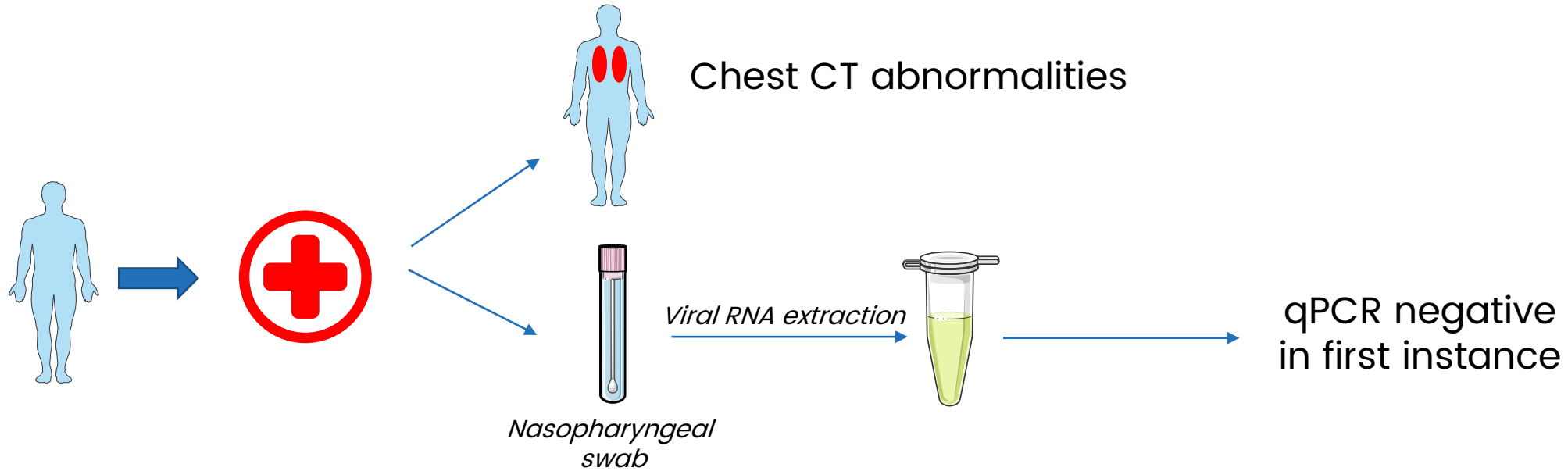


- The Crystal Digital PCR kit for COVID-19 detection was shown to reliably identify the viral sequences
 - ORF1ab: down to 5 copies per 25 μl reaction (equivalent to 0.2 cp/ μl)
 - Nucleoprotein N: down to 7 copies per 25 μl reaction (equivalent to 0.28 cp/ μl)
- No false positives were observed in 15 negative controls containing 1 ng of human RNA per 25 μl reaction

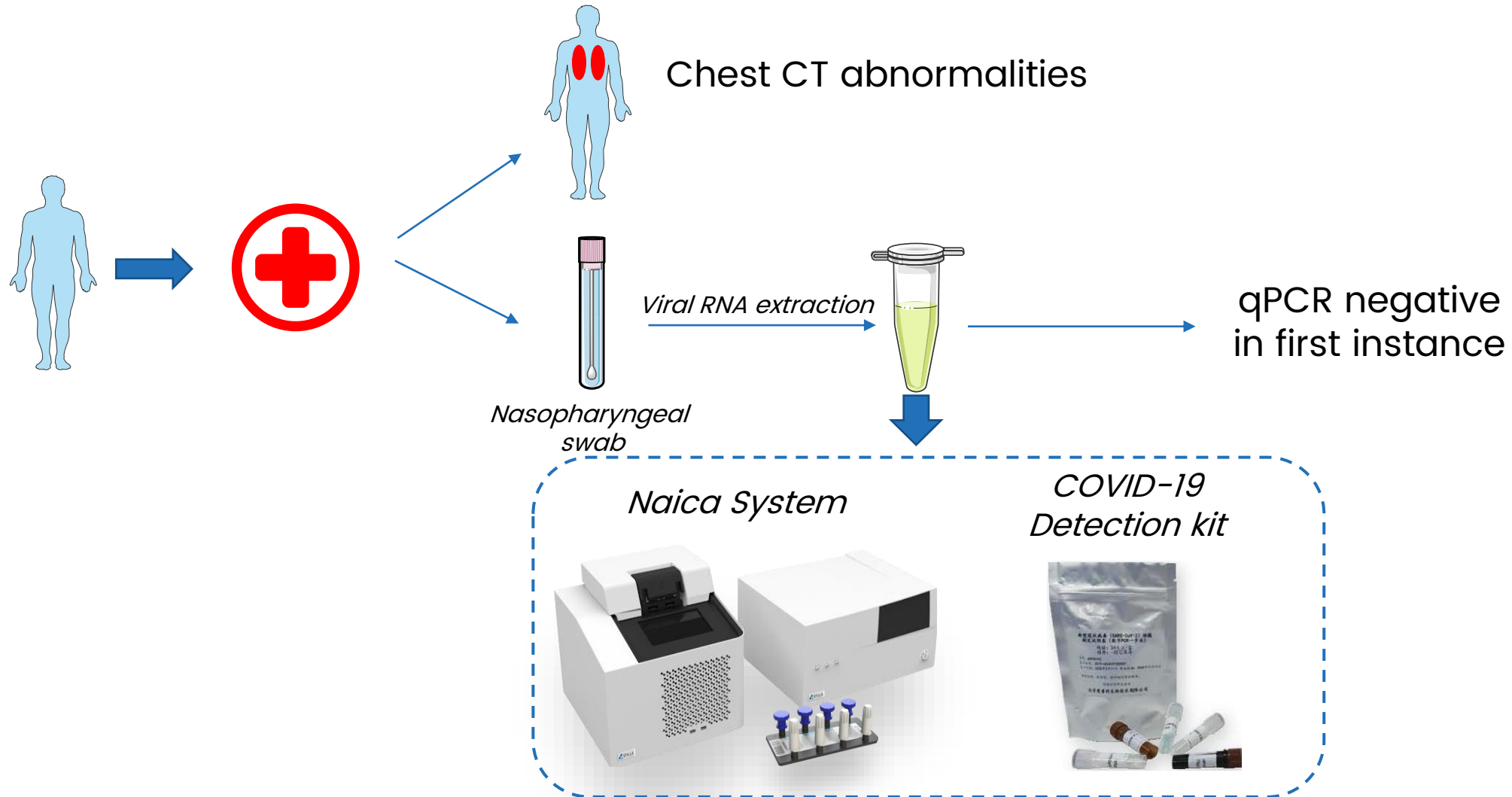


Preliminary results

Investigation of CT+/qPCR- patients



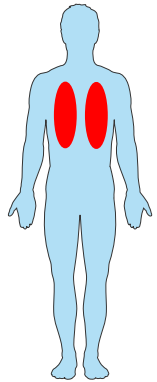
Preliminary results Investigation of CT+/qPCR- patients



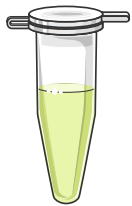
Preliminary results

Investigation of CT+/qPCR- patients

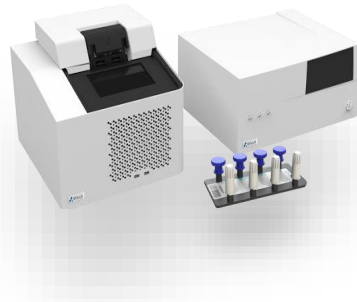
10 patients



Viral RNA extraction



qPCR negative
in first instance



| Patient # | Viral charge in Crystal Digital PCR |
|------------------|-------------------------------------|
| Patient 1 | negative |
| Patient 2 | positive |
| Patient 3 | positive |
| Patient 4 | negative |
| Patient 5 | negative |
| Patient 6 | negative |
| Patient 7 | negative |
| Patient 8 | positive |
| Patient 9 | positive |
| Patient 10 | negative |

4 patients on 10 were
retrieved positives

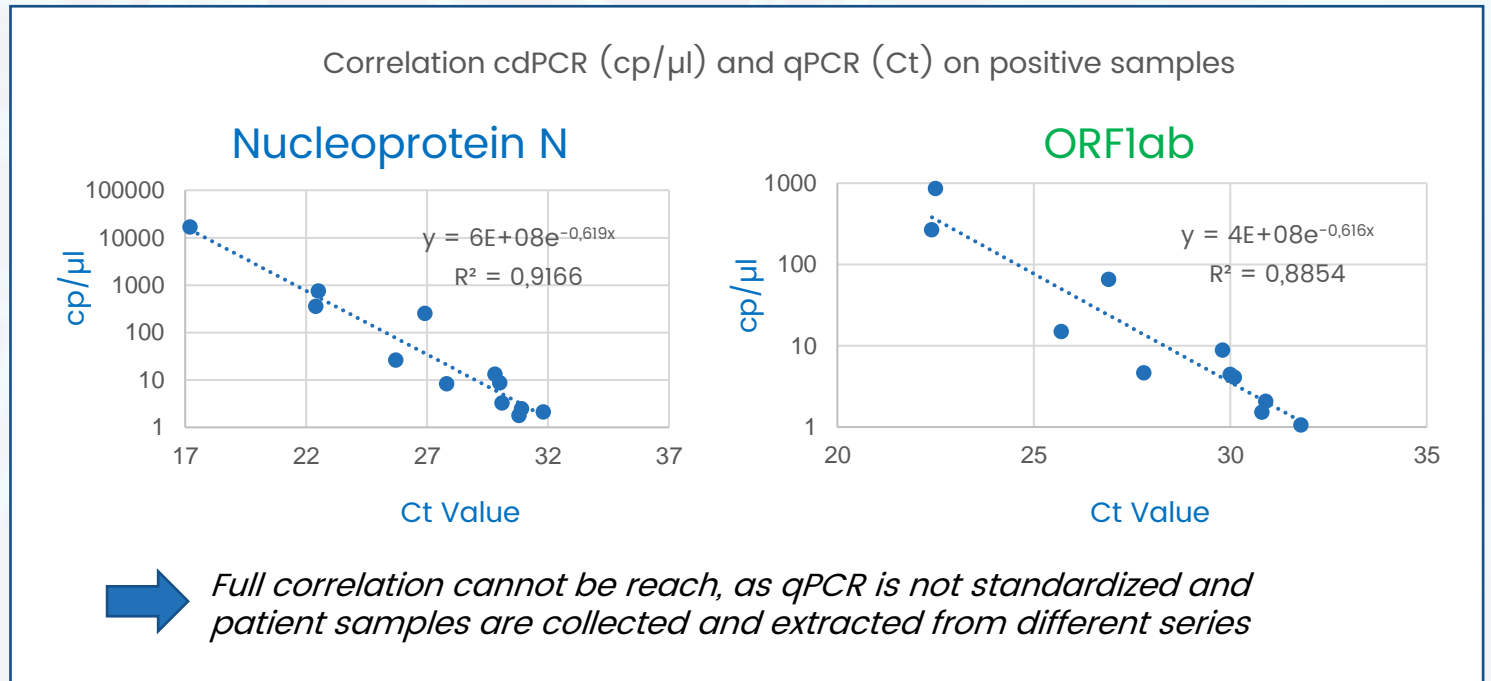
Could bring relevant
clinical information at the
first onset of the infection

COVID-19 quantification by cdPCR

- Patient samples originally tested by qPCR were evaluated with cdPCR
 - Investigate high Ct values obtained by qPCR

| qPCR | Number of patient tested |
|--|--------------------------|
| Negative <i>(Undetermined)</i> | 15 |
| Positive (Ct<35) | 12 |
| Doubtful (Ct>34) | 18 |

- One recall, determined positive by cdPCR



COVID-19 quantification by cdPCR in high Ct qPCR data

| qPCR | Number of patient tested |
|------------------------------------|--------------------------|
| Negative (Undetermined) | 15 |
| Positive (Ct<35) | 12 |
| Doubtful (Ct>34) | 18 |

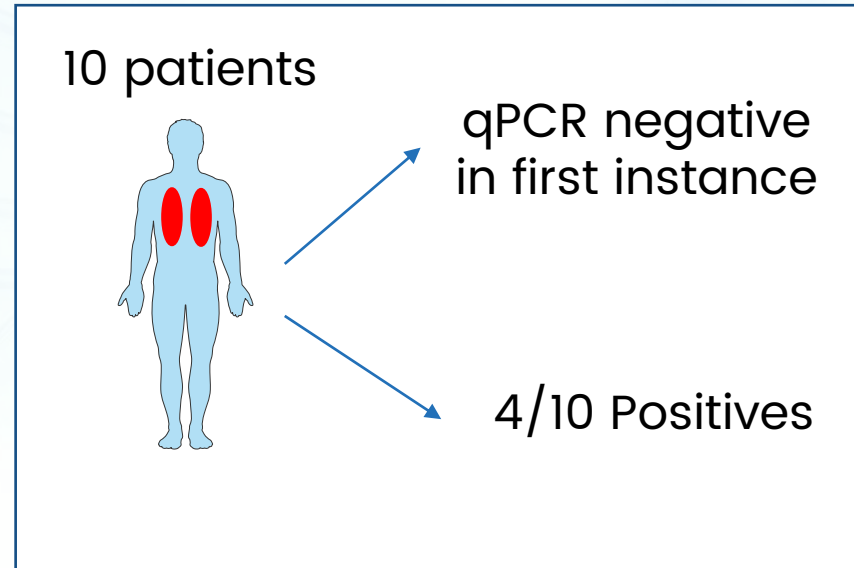
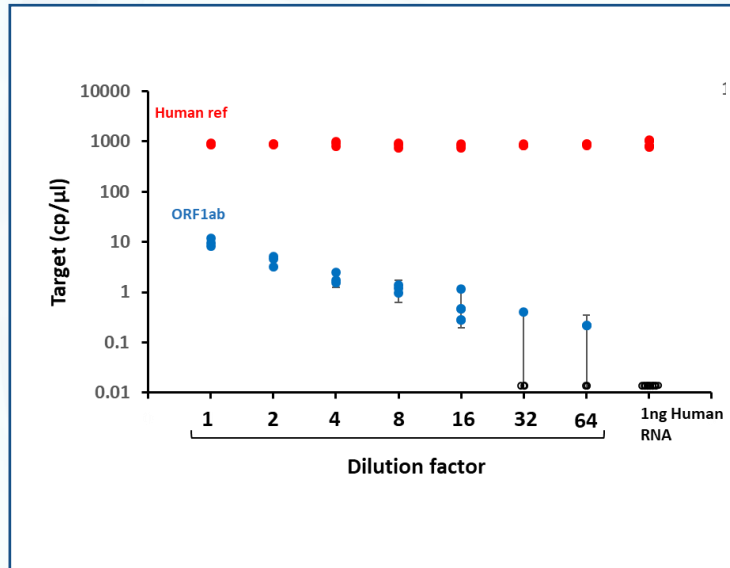
| Numéro | CT qPCR | Viral charge determined in cdPCR |
|--------|---------|----------------------------------|
| 1 | 37.32 | neg |
| 2 | 35.53 | pos |
| 3 | 35.15 | pos |
| 4 | 34.65 | neg |
| 5 | 36.92 | neg |
| 6 | 36.56 | neg |
| 7 | 36.75 | pos |
| 8 | 35.17 | Pos |
| 9 | 34.37 | pos |
| 10 | 37.43 | pos |
| 11 | 34.95 | pos |
| 12 | 38.17 | neg* |
| 13 | 33.78 | pos |
| 14 | 36,35 | pos |
| 15 | 36,66 | neg |
| 16 | 34,3 | pos |
| 17 | 36,96 | neg |
| 18 | 36,47 | pos |

**IC highlight a possible Collection/Extraction issue*

| | |
|-----------------------------------|--------------|
| Viral absence confirmed by cdPCR | 7/18 |
| Viral presence confirmed by cdPCR | 11/18 |

Crystal Digital PCR Covid-19 detection kit is a easy and fast solution to investigate difficult to interpret high Ct value qPCR data.

Conclusion



High qPCR Ct values

| | |
|-----------------------------------|-------|
| Viral absence confirmed by cdPCR | 7/18 |
| Viral presence confirmed by cdPCR | 11/18 |

- **Crystal Digital PCR™ Covid-19 detection kit can detect only few copies of the SARS-CoV2**
- **Could bring relevant clinical information at the first onset of the infection**
- **Crystal Digital PCR Covid-19 detection kit is a easy and fast solution to investigate difficult to interpret high Ct value qPCR data.**



SPECIAL THANK YOU TO:

Pr. M.Drancourt
Dr. A.Bouam

Dr. V.Thibault
Dr. C.Grolhier
Dr. M.D.Galibert
Dr. A.Lespagnol

**THANK YOU FOR YOUR
ATTENTION!
ANY QUESTIONS?**

For more information on product and workflow, visit our website at

www.stillatechnologies.com



Evaluation of Crystal Digital PCR COVID-19 detection kit by the Institut Pasteur

Reagents / Evaluations

List of diagnostic reagents by RT-PCR of SARS-CoV-2 marked CE Point on 05/05/2020 on <https://solidarites-sante.gouv.fr/IMG/pdf/liste-reactifs-diagnostic-rt-pcr.pdf>

List of CE marked COVID-19 reagents available worldwide : pdf format to download (dated 03/04/2020) here or on https://www.finddx.org/covid-19/pipeline/?avance=all&type=all&status=CE-IVD§ion=immunoassays#diag_tab

Development of the CNR on the taking of samples and the sensitivity of RT-PCR tests for the detection of SARS-CoV-2 (05/09/2020)

Inventory of kits / reagents evaluations by CNR Lyon / IP respiratory viruses (version of 05/14/2020)

Institut Pasteur reports on:

- Thermofisher V1 kit
- Thermofisher V2 kit
- R-Biopharm kit (RIDA GENE)
- NOVACYT kit (genesis)
- Eurofins Biomnis kit
- Amplidiag kit - Mobidiag
- Genefirst kit - PrimaDiag
- Bio-T SARS-CoV-2 kit - Biosellal
- Bio-T Covid-19 kit - Biosellal
- ILAMP Novel kit - Ionebio
- Lyra® kit - Quidel corporation
- IDNCOV-2 kit - idSOLUTIONS
- OPTI kit - OPTIMedical
- EXCILONE SARL kit
- BIOMAXIMA kit
- FosunPharma Diagnostics kit
- IDVet kit
- Novodiag kit - Mobidiag
- Eurobio Scientific kit
- Q-sets kit 2019 - Mediane France
- Multiplex Digital kit - Stilla
- SmartAmip kit - EMG / MSE
- IDNCOV-2d kit (s) - Id Solutions
- Tristar T-Kit Bio kit - Biosellal
- RealAccurate Quadruplex kit - AirDiag_Pathfinder

Lyon reports on:

- Xpert® kit - Cepheid
- CerTest kit - BD
- BIOGX kit - Launch Diagnostics
- ARGENE kit - bioMérieux
- COBAS kit - Roche Diagnostics
- VitaPCR™ kit - BioSynex
- RealStar®SARS-CoV-2 kit - AltonaDiagnostic
- BD and R-Biopharm coupled kit
- VIASURE kit - Orgentec
- BioFire® kit - bioMérieux on FilmArray®Torch
- Bosphore® kit - Launch Diagnostics
- Combo2Screen kit - ABI
- kit Simplexa™ - Diasorin Molecular
- Sansure Biotech kit

■ Multiplex Digital kit - Stilla

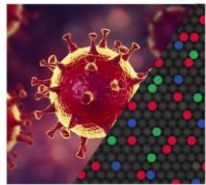
Report available on:

<https://www.sfm-microbiologie.org/2020/05/11/covid-19/>

APPLICATION & TECHNICAL NOTES



A 3-color Crystal Digital PCR™ kit for detection of COVID-19



Development of one-step RT-dPCR models for COVID-19 detection

The 2019-2020 outbreak of COVID-19 caused by the SARS-CoV-2 virus first reported in Wuhan, Hubei, China has been declared a pandemic by the World Health Organization. To facilitate the action of health authorities, the development of robust laboratory tests is of primary importance. Using the numerous publicly accessible SARS-CoV-2 and SARS-related sequences, several PCR-based assays specific for SARS-CoV-2 have been designed (Chan et al., 2020). The Naica compatible 3-color Crystal Digital PCR™ kit (Figure 1), developed by ApexBio (Hsinchu Science-based Industrial Park) includes primers and FAM- and HEX-labeled probes specific to two distinct regions (ORF1ab and Nucleocapside (N) genes, respectively) of the SARS-CoV-2 positive strand RNA genome. The 3rd channel of the Naica™ system has been used as an endogenous PCR reference detecting a human housekeeping gene with a Cy5-labeled probe. This single assay design permits the simultaneous detection of two independent SARS-CoV-2 sequences reported as conserved while concurrently monitoring PCR effectiveness using the third channel of detection (Figure 2).



Kit components

- dPCR master mix1
- dPCR master mix2
- Primers and probes mix
- SARS-CoV-2 positive control
- SARS-CoV-2 negative control

Figure 1: The RUO ApexBio-developed ready-to-use kit contains all reagents required to perform a one-step RT 3-color Crystal Digital PCR™ on the Naica™ System.

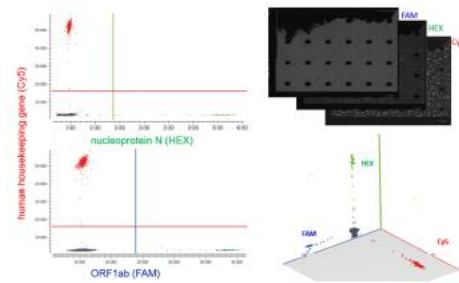


Figure 2: Crystal Miner-generated 2D (left) and 3D dot plots (right) and crystal droplet images obtained on positive controls containing human RNA and synthetic target sequences of the RUO RT-dPCR SARS-CoV-2 detection kit.

Chan JF, Yip CC, To KK, Tang TH, Wong SC, Leung KH, Fung AY, Ng AC, Zou Z, Tsui HW, Choi GK, Tam AR, Cheng VC, Chen KH, Tsang OT, Yuan KY. Improved molecular diagnosis of COVID-19 by the novel, highly sensitive and specific COVID-19-RdRp/rtRt real-time reverse transcription-polymerase chain reaction assay validated in vitro and with clinical specimens. J Clin Microbiol. 2020 Mar 4.

Sensitive and specific detection of COVID-19

An experimental model containing synthetic sequences targeted by the SARS-CoV-2 detection kit was serially diluted and seven dilution points were assessed in triplicate. A total of 1ng of human RNA was added to each replicate. The results indicated a robust and specific detection of SARS-CoV-2 sequences down to 0.8 copies per μ l of positive control (5 copies per 25 μ l reaction) of the ORF1ab gene and down to 0.9 copies per μ l of positive control (7 copies per 25 μ l reaction) of the Nucleocapside (N) gene in all tested samples. Further dilutions showed an extremely sensitive but stochastic detection down to 0.25 copies per μ l of positive control (2 copies per 25 μ l reaction) for both genes (Figure 3). In parallel, a total of 15 controls containing only human RNA were tested as negative controls and no false positives were observed.

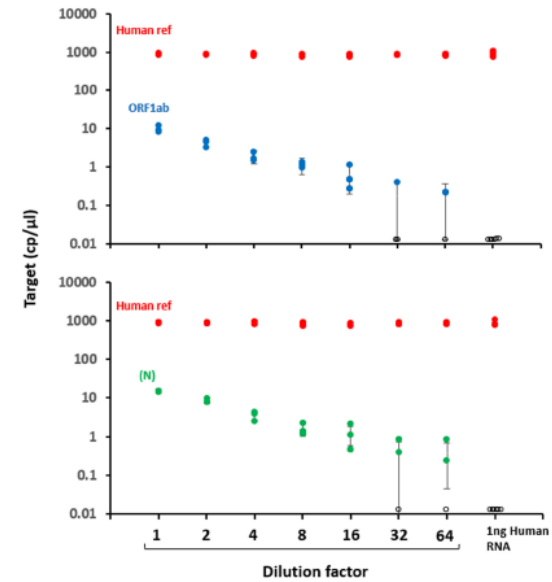
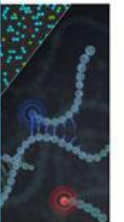


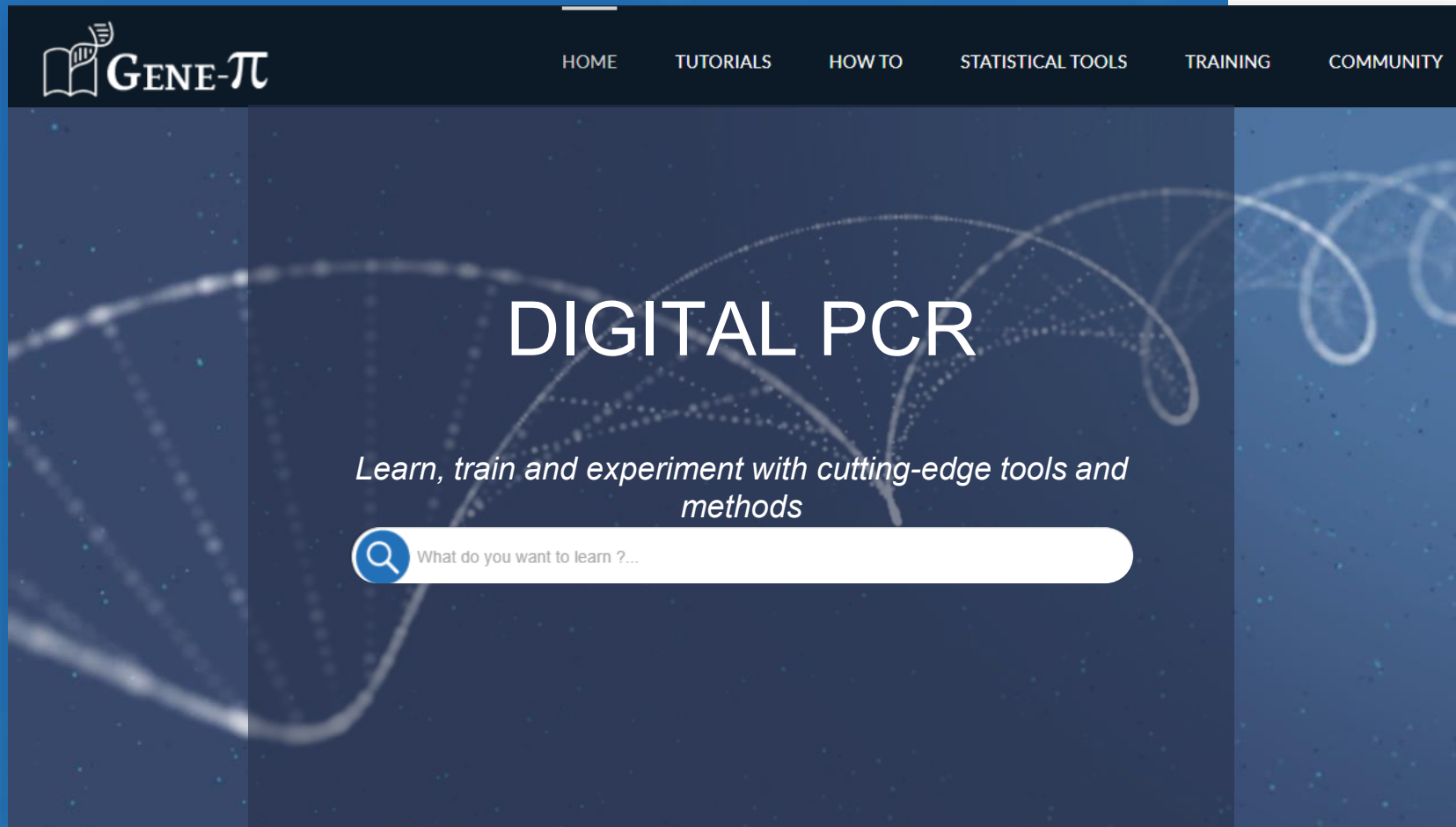
Figure 3: Sensitivity of the 3-color RUO RT Crystal Digital PCR™ kit for SARS-CoV-2 detection. Serial dilutions of SARS-CoV-2 synthetic targets were assayed in triplicate in a background of 1ng of human RNA. A total of 8 μ l of positive controls was added to each 25 μ l reaction. The vertical bars represent the theoretical 95% Poisson confidence intervals for the pool of 3 replicates. The empty circles represent replicates where SARS-CoV-2 sequences were not detected.

LA
Simplify Drop-Off
PCR assays
Crystal Miner™



Crystal Digital PCR VS Quantitative PCR

| | Quantitative PCR | Digital PCR |
|----------|--|---|
| Process | <ul style="list-style-type: none">• Same Sample preparation methods and reagents• Similar initial sample volumes• Capability of multiplexing (amplifying several different DNA sequences simultaneously) | |
| Analysis | <ul style="list-style-type: none">• Standard curve required | <ul style="list-style-type: none">• No standard curve required |
| Results | <ul style="list-style-type: none">• Relative quantification• Reproducible results rely on human expertise | <ul style="list-style-type: none">• Absolute quantification• Lower variability |
| Usage | <ul style="list-style-type: none">• Monitoring of real-time reaction efficiency• Relative gene expression if differences are >2-fold | <ul style="list-style-type: none">• High sensitivity and reproducibility• Rare allele detection• Easy to use |



LAUNCH IN MARCH 2019:



3 tutorials

- Rare Mutation Detection
- CNV
- Drop-off Assay



1 video



14 how to's



3 memos



3 online statistical tools

- Poisson Law
- CNV
- Limit of Blank/Limit of Detection

