



# CRYSTAL DIGITAL PCR™ DETECTION OF SARS-CoV-2 USING THE NAICA™ SYSTEM

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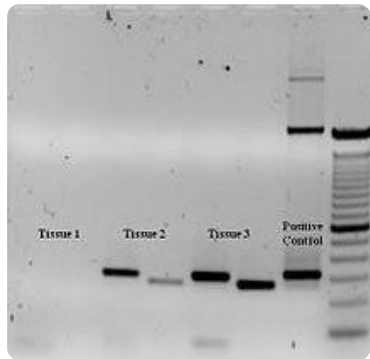
Kimberley Gutierrez, PhD  
Senior Field Application Scientist  
Stilla Technologies Inc.

# NEXT GENERATION OF PCR

## Digital PCR

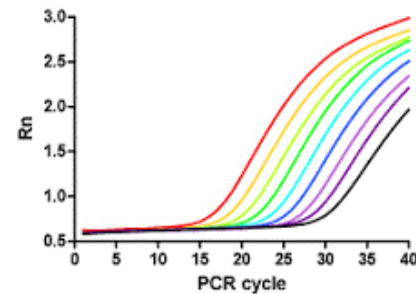
OUR MISSION:  
MAKE DIGITAL PCR A LAB COMMODITY

### PCR



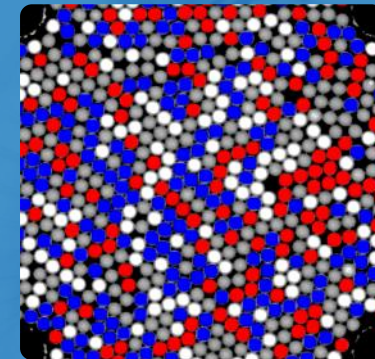
Amplify Target DNA

### Quantitative PCR



Relative quantification  
Real-time with standard curves  
Ubiquitously spread method

### Digital PCR

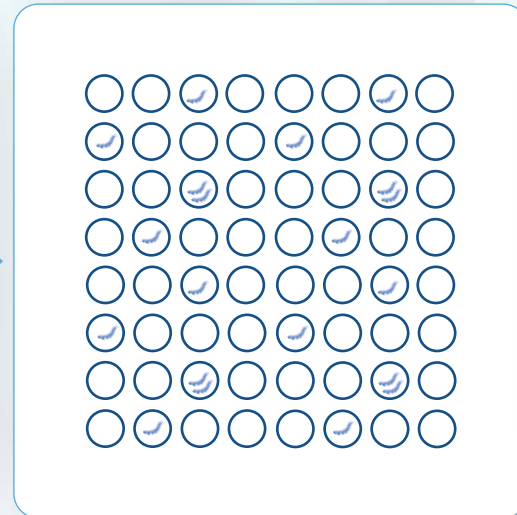
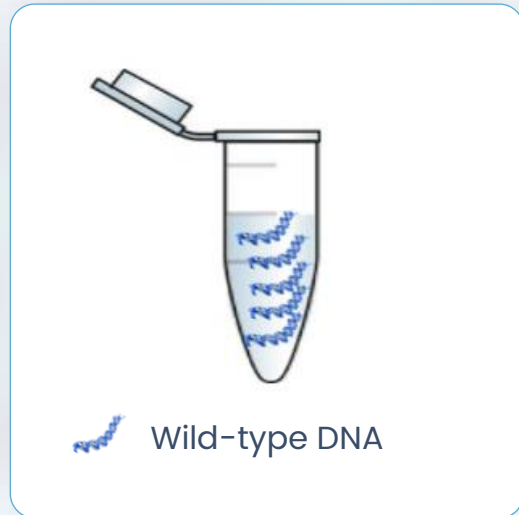


Absolute quantification  
No standard curve  
(endpoint PCR)  
Increased sensitivity

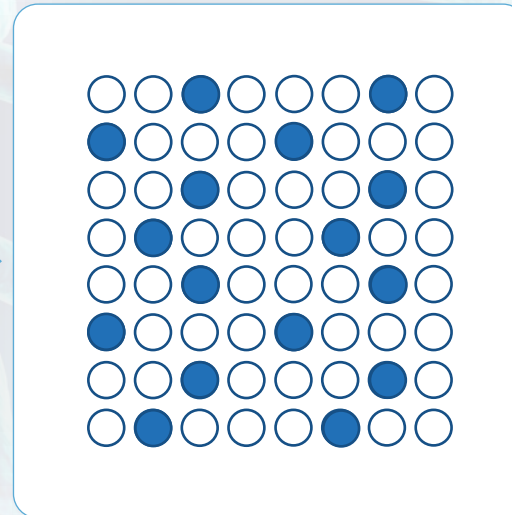


# PRINCIPLE OF DIGITAL PCR

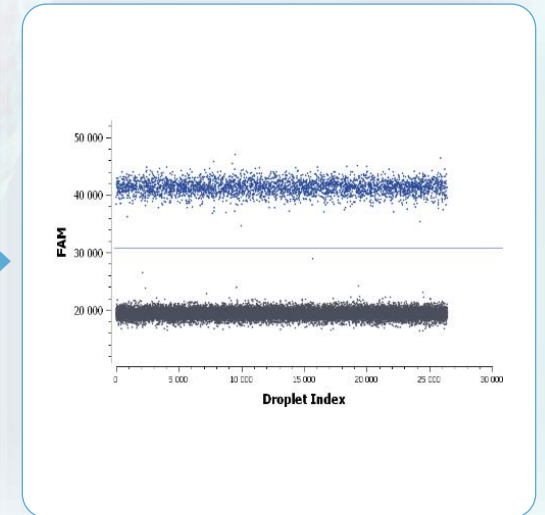
## PARTITIONING



## PCR



## READING & ANALYSIS



**RESULTS**  
2636 cp/μL with 2.2 %  
uncertainty

POISSON STATISTICS

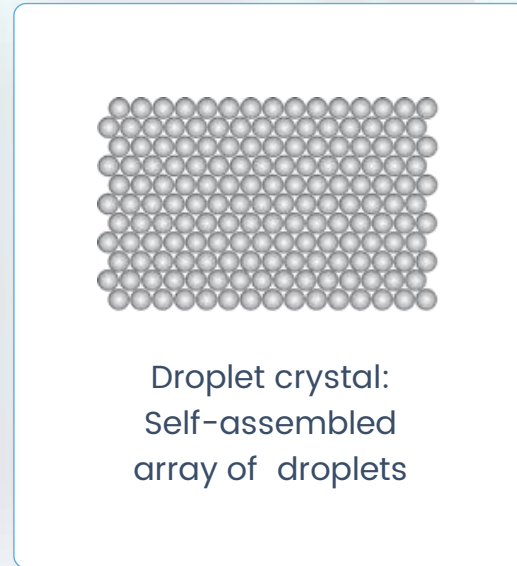
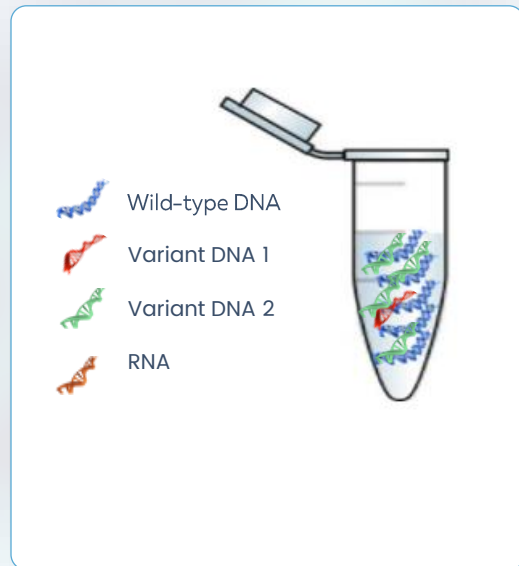
$$\frac{N_{pos}}{N_{tot}}$$



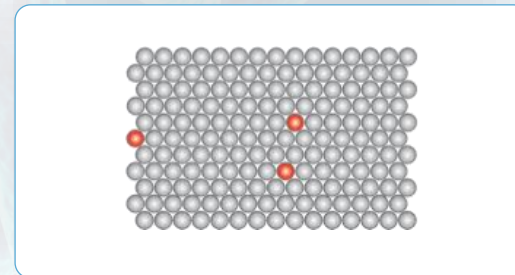
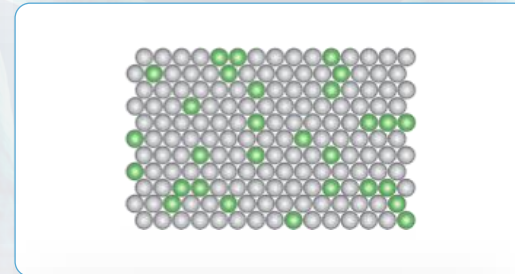
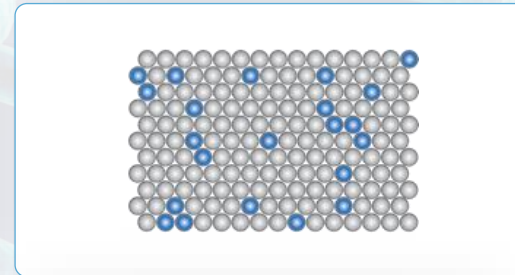


# PRINCIPLE OF CRYSTAL DIGITAL PCR™

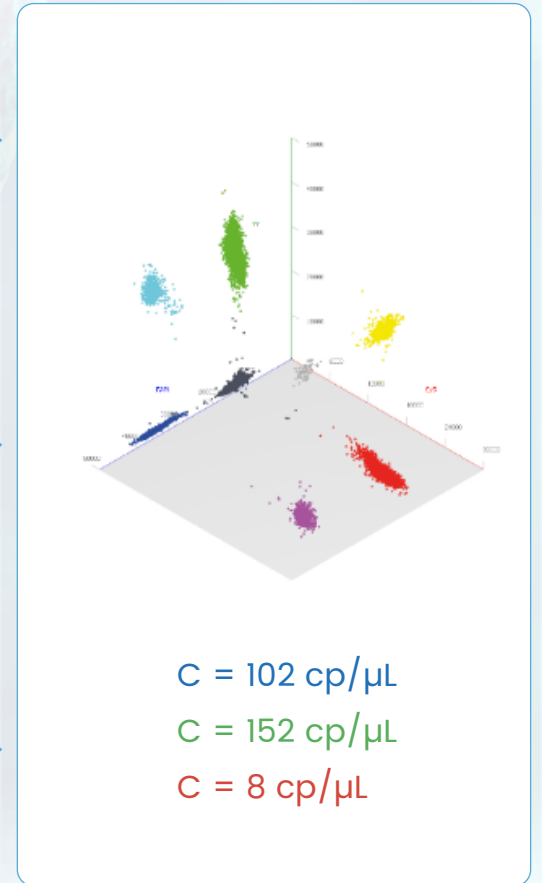
## PARTITIONING



## PCR



## READING & ANALYSIS



### 2 Parameters for accurate quantification in dPCR:

- Number of droplets
- Size of the droplets



# THE UNIQUE FEATURES OF THE NAICA™ SYSTEM



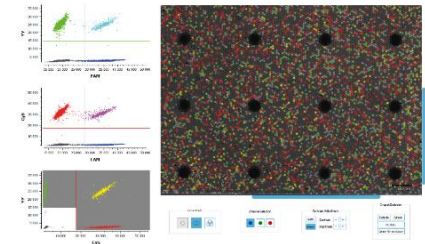
**Sapphire Chip**  
(consumable)



**Naica Geode**



**Naica Prism3**



**Crystal Miner**  
(software)



**An easy-to-use and  
integrated solution  
for digital PCR**



**Fast time-to-result  
(2h30)**



**Reliable multiplex  
assays with  
3-color detection**

*“We are extremely satisfied with the Naica System, which fully answers our needs in terms of precision and reproducibility for liquid biopsy testing.”*

**Dr. Ludovic LACROIX**

Dir. Translational Research /  
Institut Gustave Roussy



# AT THE HEART OF OUR INNOVATION: THE SAPPHIRE CHIP

A patented partitioning technology: **droplet crystals**.

## Sapphire Chip pre-filled with oil

Input volume	<b>25 <math>\mu</math>L</b>
Droplets per sample	<b>~ 30 000</b>
Droplet volume	<b>0.59 nL</b>
Number of samples	<b>4 / chip</b>
LOD	<b>0.2 cp/<math>\mu</math>L</b>
Dynamic range	<b>5 logs</b>



### Droplet crystal:

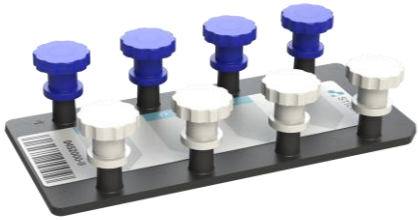
Self-assembled  
array of droplets

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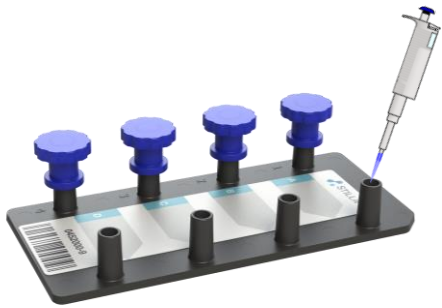


## STEP 1

# PREPARE THE SAPPHIRE CHIP – 5 MIN



UNPACK SAPPHIRE CHIP



PIPETTE 25  $\mu$ L OF PCR MIX



SEAL INLET PORT WITH CAP

## COMPATIBLE MIXES AND CHEMISTRIES:

Use with Quanta BioSciences PCR and RT-PCR Mix with no ROX

With TaqMan™ Probes, add Fluorescein as reference dye





## STEP 2 PARTITION & AMPLIFY – 2H10



### STEP 2.1 – PARTITION

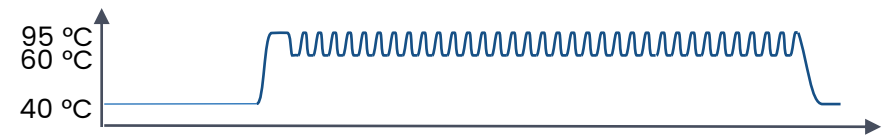
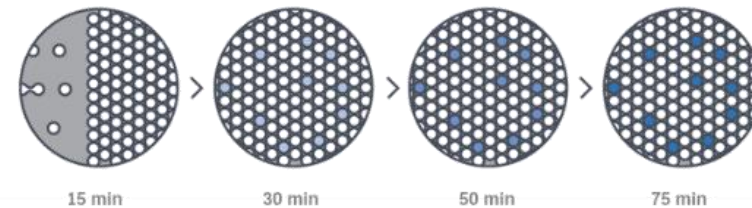
#### Charge chips into the Geode

- 1-3 chips and 1-12 samples/run
- ~30,000 partitions/sample

#### Contactless fluid injection

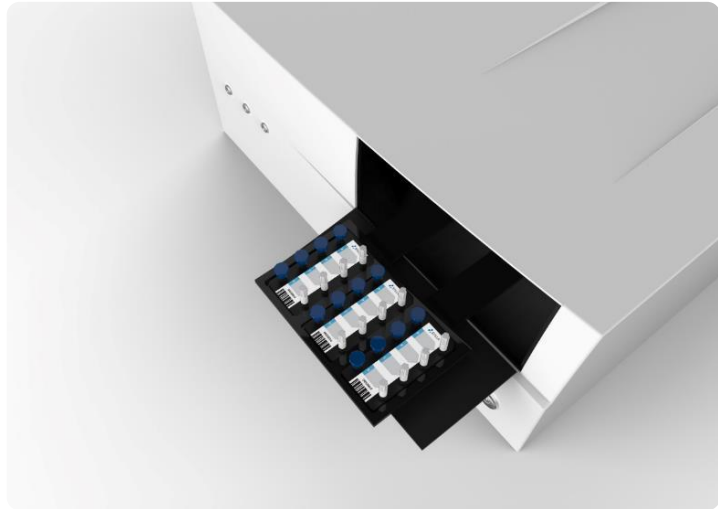
### STEP 2.2 – AMPLIFY

#### Standard cycling conditions:



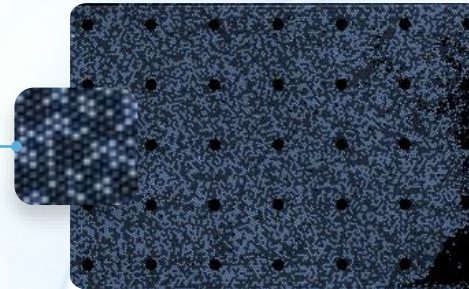


## STEP 3 DETECT – 10 MIN (50s/SAMPLE)



### TRANSFER CHIPS TO THE PRISM3

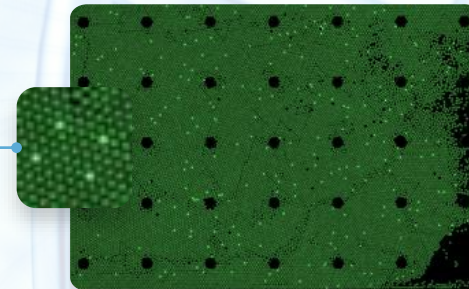
- 1-3 chips and 1-12 samples/run
- 3 color fluorescence imaging



#### **Blue**

Ex: 415-480 nm  
Em: 495-520 nm

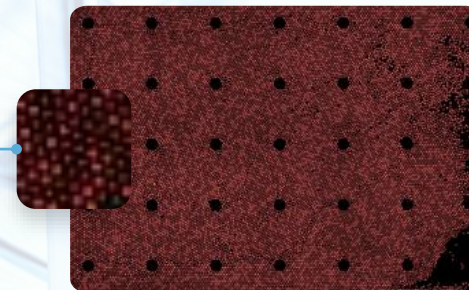
FAM...



#### **Green**

Ex: 530-550 nm  
Em: 560-610 nm

ROX, HEX, Cy3...



#### **Red**

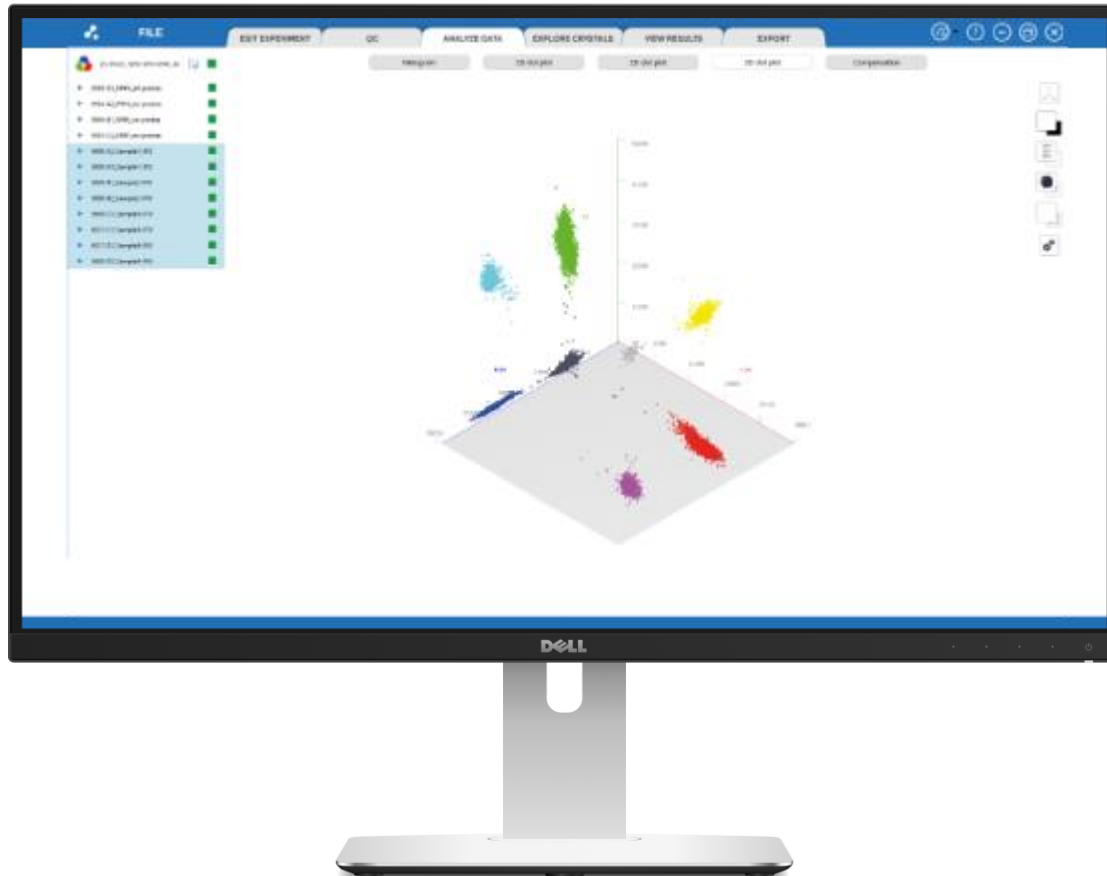
Ex: 615-645 nm  
Em: 655-720 nm

Cy5, Cy5.5...



## STEP 4

# ANALYZE YOUR DATA WITH THE CRYSTAL MINER SOFTWARE



User-friendly software with intuitive visuals



Simple image analysis and data exploration



A trustworthy quality control



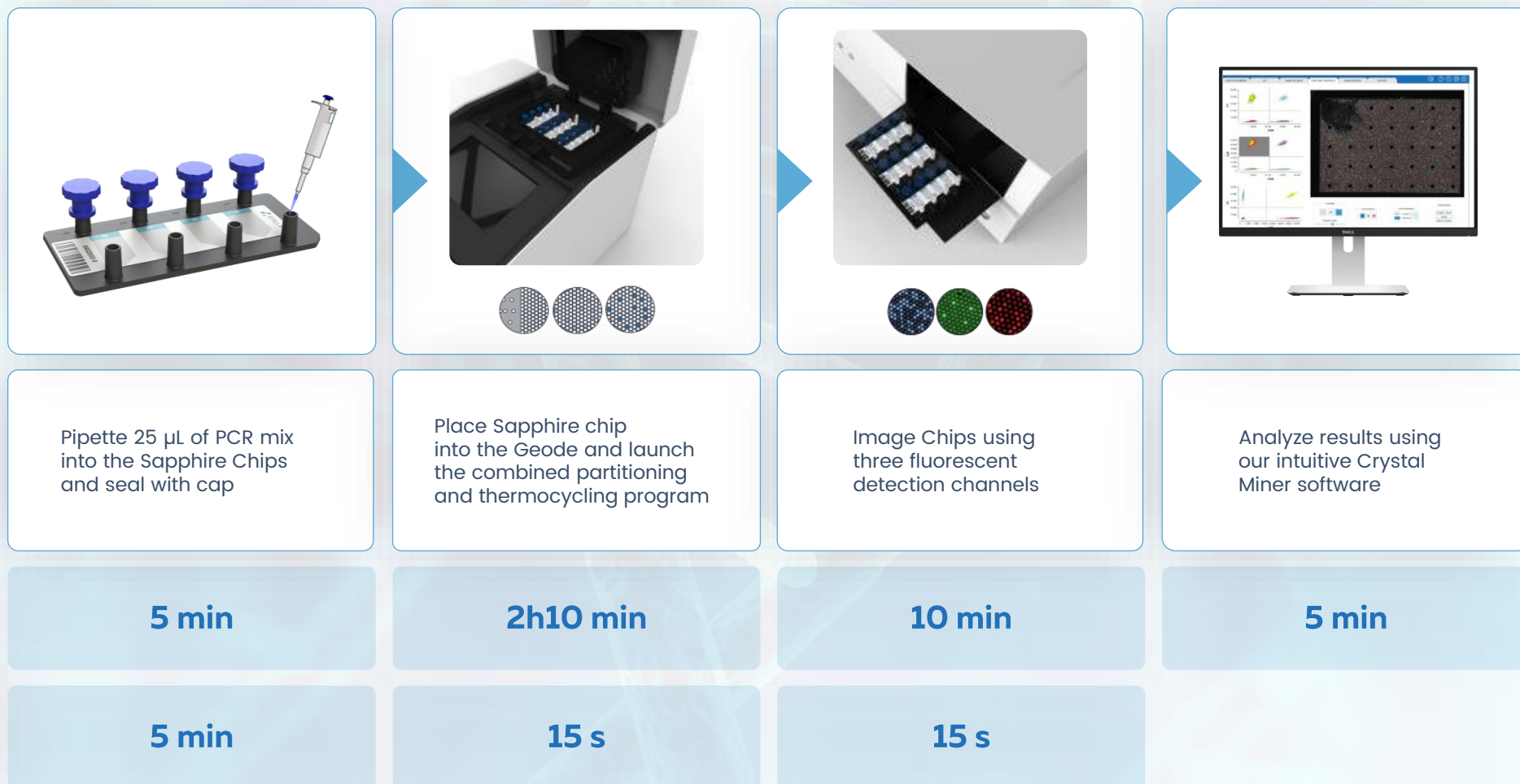
# EXPLORE CRYSTALS

The screenshot displays the CrystalMiner software interface. At the top, there are menu options: FILE, QUALITY CONTROL, SETUP, ANALYZE DATA (with sub-options Plots & Populations and Explore Crystals), VIEW RESULTS, and EXPORT. Below the menu is a list of 16 samples, with the 8th sample, '04326717-D1\_556', selected. To the right of the list are three flow cytometry plots. The top plot shows HEX (pUC18 MCS L1) vs FAM ( BRAF WT) with a green gate. The middle plot shows Cy5 ( ALB) vs FAM ( BRAF WT) with a red gate. The bottom plot shows HEX (pUC18 MCS L1) vs Cy5 ( ALB) with a yellow gate. To the right of the plots is a large image of a crystal array with a 1.8 mm scale bar. Below the image are four control panels: Color Mode (with a Population opacity slider), Channel Selection (with blue, green, and red buttons), Contrast Adjustment (with Auto, Contrast, Brightness, and Reset buttons), and Droplet Exclusion (with Exclude, Cancel, Restore, and Select for exclusion buttons).

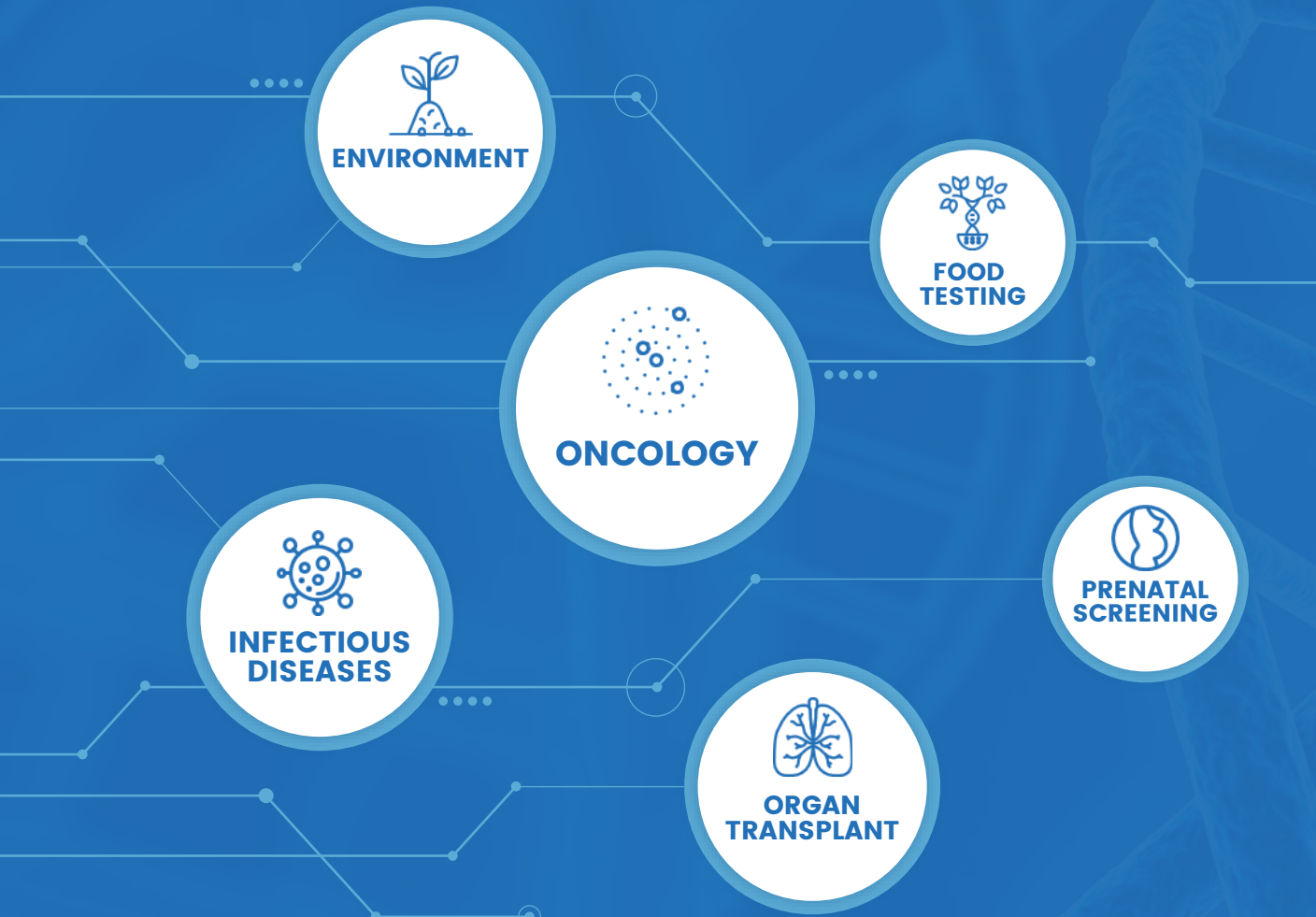




# PERFORM CRYSTAL DIGITAL PCR™ IN 2H30 WITH MINIMUM HANDS-ON TIME



# POTENTIAL APPLICATIONS



**+3300**  
publications for digital PCR in 2018\*  
Oncology is driving the field

- TYPE OF ASSAYS:**
- ✓ Absolute quantification (DNA/RNA)
  - ✓ Copy number variation
  - ✓ Rare event detection
  - ✓ Gene expression

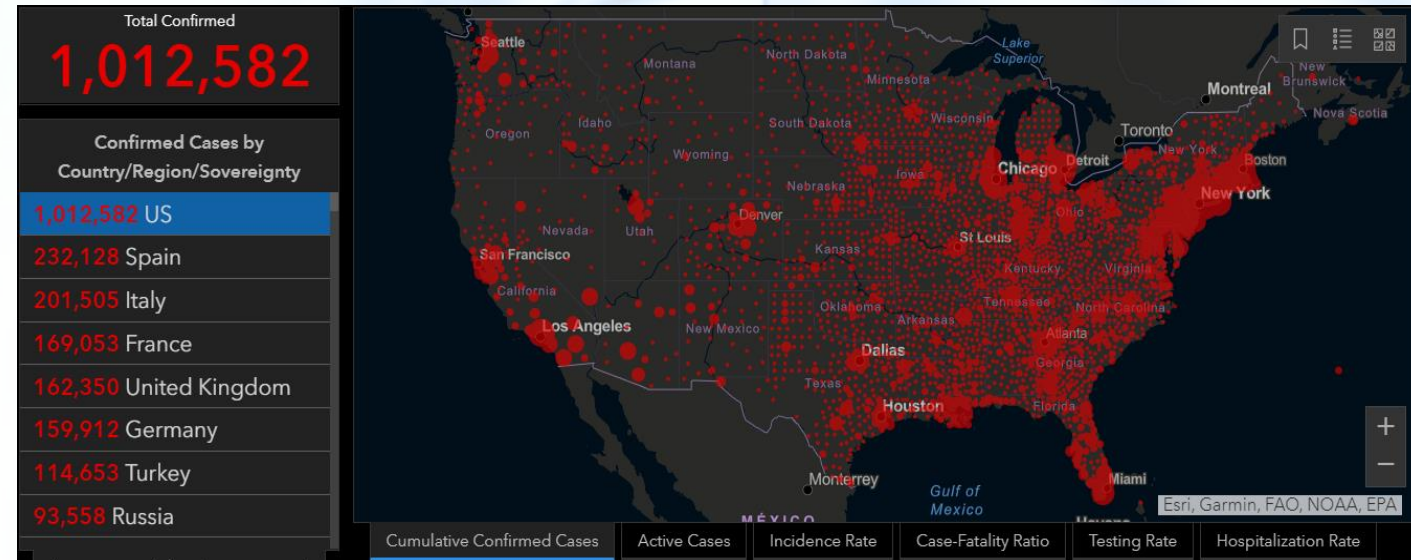
\*Source: Google Scholar



# COVID-19 PANDEMIC SARS-CoV-2

As of April 28, 2020:

- More than 3 million cases reported worldwide
  - 185 countries and regions
- More than 1 million cases confirmed in the US
- More than 5.7 million tests for COVID-19 have been conducted in the US
  - 1.7% of the US population tested



Need for more research to understand the clinical outcomes of infection

**Stilla has partnered with ApexBio to offer a digital PCR kit to detect SARS-CoV-2**



# A novel RUO kit for COVID-19 detection in human samples *developed by ApexBio*

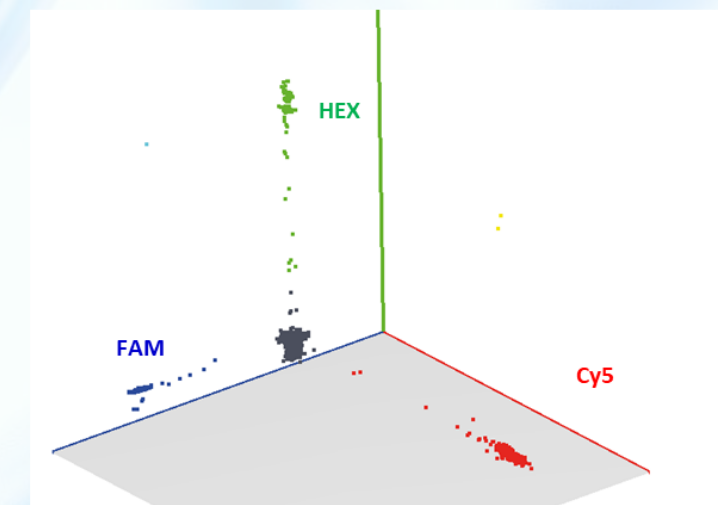
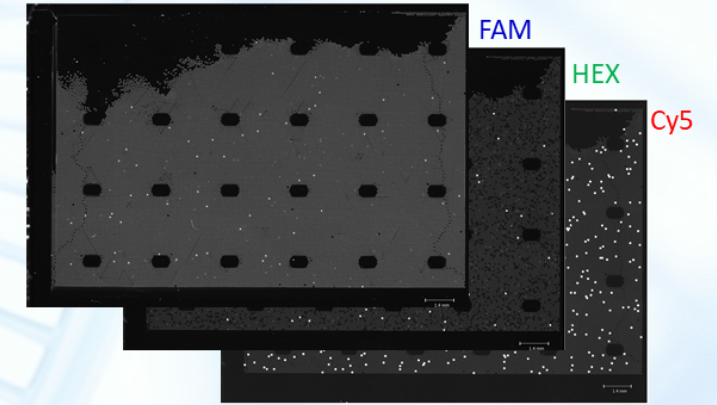
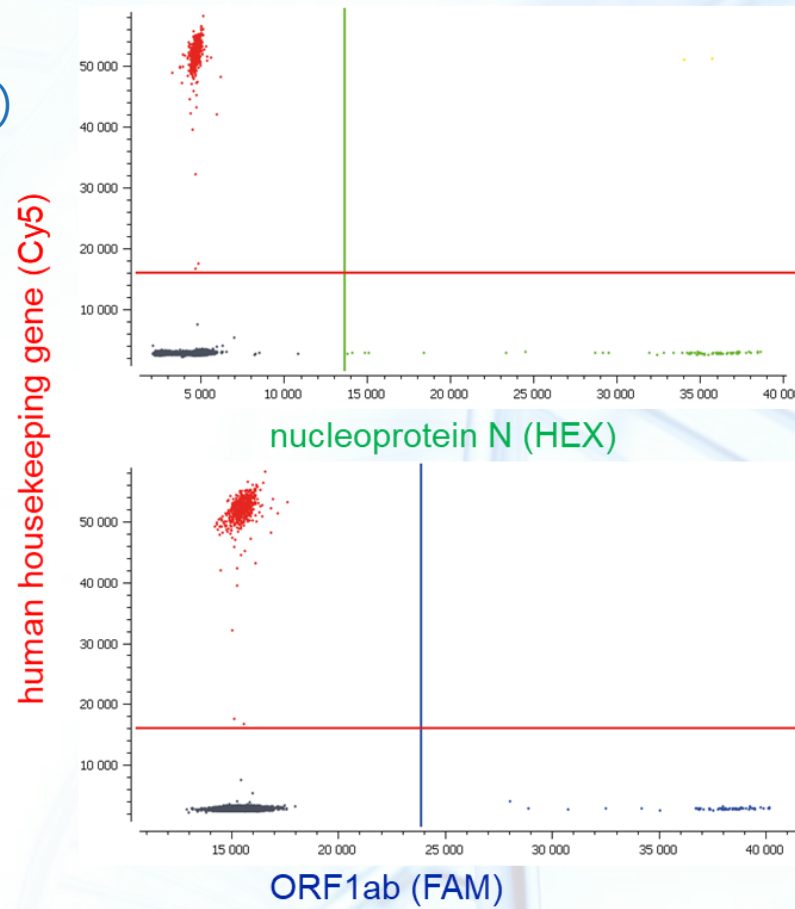
## 3-color kit to detect viral and human genes:

- COVID-19 ORF1ab (FAM)
- COVID-19 nucleoprotein N (HEX)
- Human control housekeeping gene (Cy5)



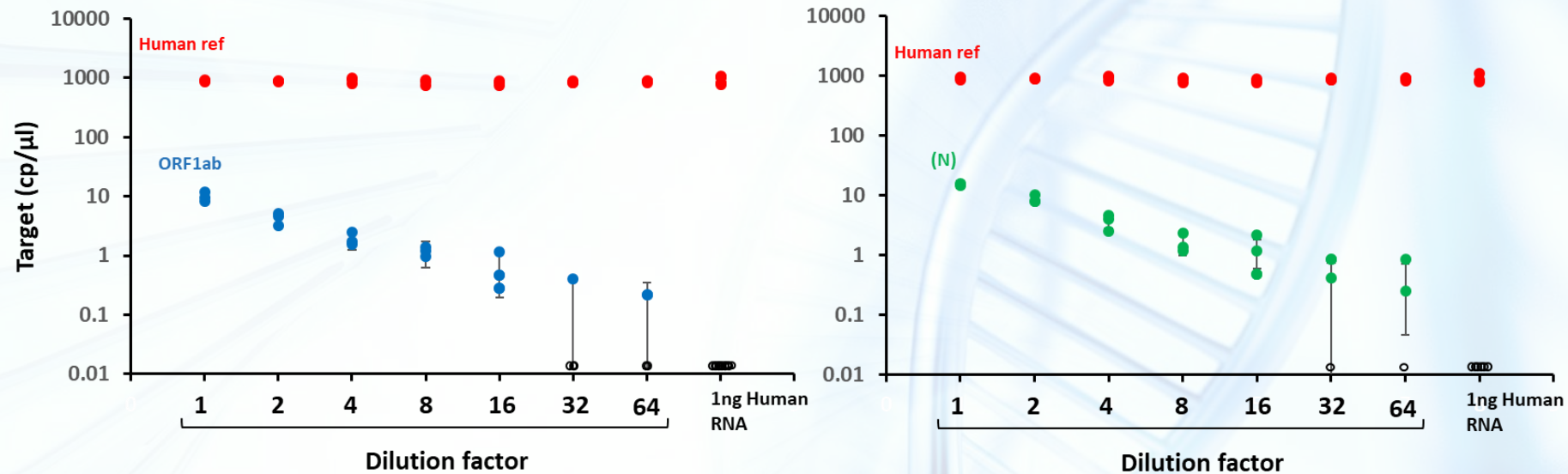
## Kit components

- dPCR master mix1
- dPCR master mix2
- Primer and probe mix
- COVID-19 positive control
- COVID-19 negative control



# Sensitive and specific detection of COVID-19 sequences

- A positive control containing ORF1ab and nucleoprotein N sequences was serially diluted and tested in triplicate.
- A total of 8  $\mu$ l of positive control was assessed in a 25 $\mu$ l reaction in a background of 1 ng of human RNA.



- The Crystal Digital PCR kit for COVID-19 detection was shown to reliably identify the viral sequences
  - 0.6 cp/ul of ORF1ab (equivalent to 5 copies per 25ul reaction)
  - 0.9 cp/ul of nucleoprotein N (equivalent to 7 copies per 25 $\mu$ l reaction)
- No false positives were observed in 15 negative controls containing 1 ng of human RNA per 25  $\mu$ l reaction

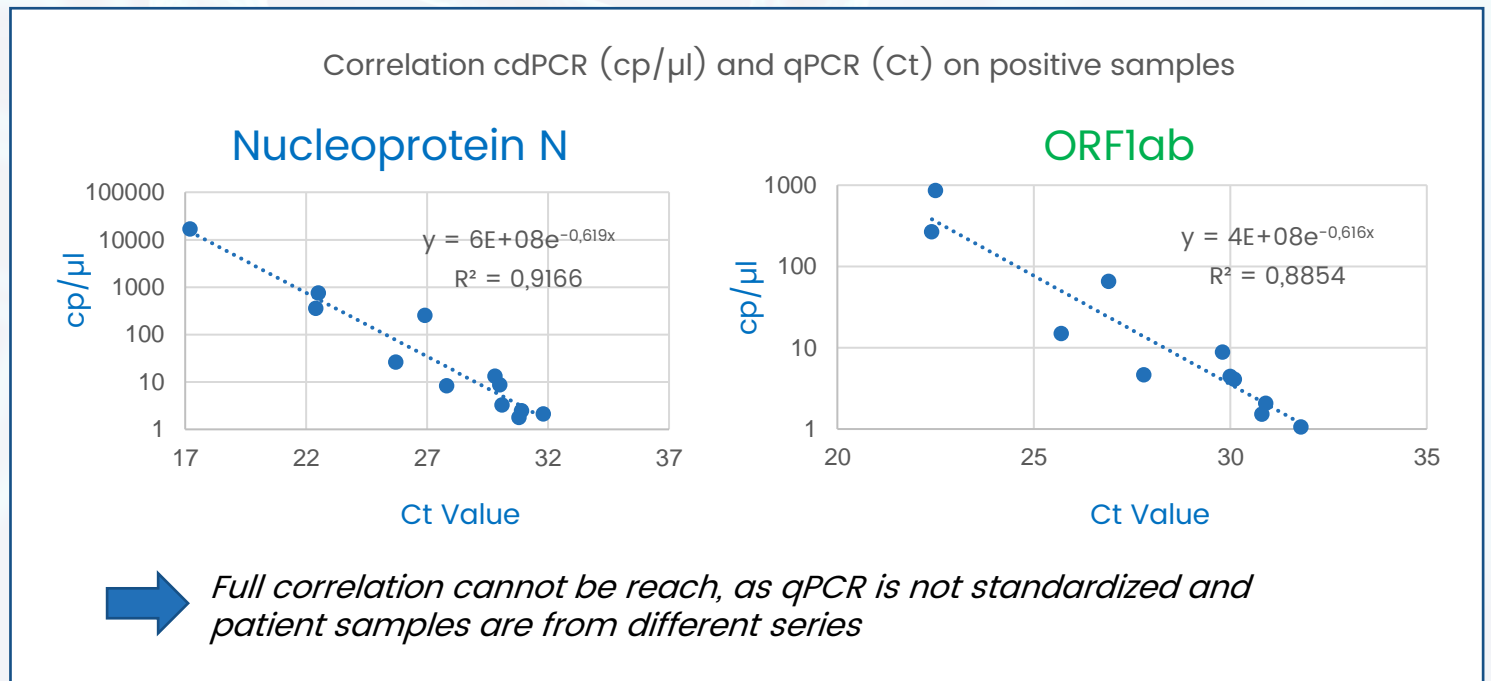


# COVID-19 quantification by cdPCR

- Patient samples originally tested by qPCR were revalidated with cdPCR
  - Investigate high Ct values obtained by qPCR

qPCR	Number of patient tested
<b>Negative</b> <i>(Undetermined)</i>	<b>15</b>
<b>Positive (Ct&lt;35)</b>	<b>12</b>
<b>Doubtful (Ct&gt;34)</b>	<b>18</b>

- One recall, determined positive by cdPCR





# COVID-19 quantification by cdPCR in high Ct qPCR data

qPCR	Number of patient tested
<b>Negative (Undetermined)</b>	<b>15</b>
<b>Positive (Ct&lt;35)</b>	<b>12</b>
<b>Doubtful (Ct&gt;34)</b>	<b>18</b>

Number	CT qPCR	Viral designation determined in cdPCR
1	37.32	neg
2	35.53	pos
3	35.15	pos
4	34.65	neg
5	36.92	neg
6	36.56	neg
7	36.75	pos
8	35.17	Pos
9	34.37	pos
10	37.43	pos
11	34.95	pos
12	38.17	neg*
13	33.78	pos
14	36,35	pos
15	36,66	neg
16	34,3	pos
17	36,96	neg
18	36,47	pos

*\*IC highlight a possible Collection/Extraction issue*

Viral absence confirmed by cdPCR	<b>7/18</b>
Viral presence confirmed by cdPCR	<b>11/18</b>

**High sensitive cdPCR Covid-19 detection kit is a powerful solution to investigate difficult to interpret high Ct value qPCR data.**

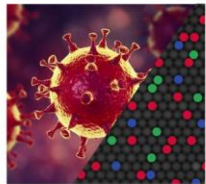
# Digital PCR VS Quantitative PCR

	Quantitative PCR	Digital PCR
Process	<ul style="list-style-type: none"><li>• Same Sample preparation methods and reagents</li><li>• Similar initial sample volumes</li><li>• Capability of multiplexing (amplifying several different DNA sequences simultaneously)</li></ul>	
Analysis	<ul style="list-style-type: none"><li>• <b>Standard curve required</b></li></ul>	<ul style="list-style-type: none"><li>• <b>No standard curve required</b></li></ul>
Results	<ul style="list-style-type: none"><li>• <b>Relative quantification</b></li><li>• <b>Reproducible results rely on human expertise</b></li></ul>	<ul style="list-style-type: none"><li>• <b>Absolute quantification</b></li><li>• <b>Lower variability</b></li></ul>
Usage	<ul style="list-style-type: none"><li>• Monitoring of real-time reaction efficiency</li><li>• Relative gene expression if differences are &gt;2-fold</li></ul>	<ul style="list-style-type: none"><li>• <b>High sensitivity and reproducibility</b></li><li>• <b>Rare</b> allele detection</li></ul>

# APPLICATION & TECHNICAL NOTES



## A 3-color Crystal Digital PCR™ kit for detection of COVID-19



### Development of one-step RT-dPCR models for COVID-19 detection

The 2019-2020 outbreak of COVID-19 caused by the SARS-CoV-2 virus first reported in Wuhan, Hubei, China has been declared a pandemic by the World Health Organization. To facilitate the action of health authorities, the development of robust laboratory tests is of primary importance. Using the numerous publicly accessible SARS-CoV-2 and SARS-related sequences, several PCR-based assays specific for SARS-CoV-2 have been designed (Chan et al., 2020). The Naica compatible 3-color Crystal Digital PCR™ kit (Figure 1), developed by ApexBio (Hsinchu Science-based Industrial Park) includes primers and FAM- and HEX-labeled probes specific to two distinct regions (ORF1ab and Nucleocapside (N) genes, respectively) of the SARS-CoV-2 positive strand RNA genome. The 3rd channel of the Naica™ system has been used as an endogenous PCR reference detecting a human housekeeping gene with a Cy5-labeled probe. This single assay design permits the simultaneous detection of two independent SARS-CoV-2 sequences reported as conserved while concurrently monitoring PCR effectiveness using the third channel of detection (Figure 2).



#### Kit components

- dPCR master mix1
- dPCR master mix2
- Primers and probes mix
- SARS-CoV-2 positive control
- SARS-CoV-2 negative control

Figure 1: The RUO ApexBio-developed ready-to-use kit contains all reagents required to perform a one-step RT 3-color Crystal Digital PCR™ on the Naica™ System.

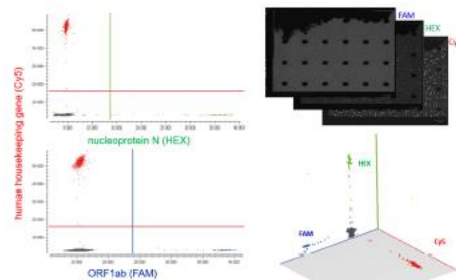


Figure 2: Crystal Miner-generated 2D (left) and 3D dot plots (right) and crystal droplet images obtained on positive controls containing human RNA and synthetic target sequences of the RUO RT-dPCR SARS-CoV-2 detection kit.

Chan JF, Yip CC, To KK, Tang TH, Wong SC, Leung KH, Fung AY, Ng AC, Zou Z, Tsui HW, Choi GK, Tam AR, Cheng VC, Chen KH, Tsang OT, Yuan KY. Improved molecular diagnosis of COVID-19 by the novel, highly sensitive and specific COVID-19-RdRp/rtRdRr real-time reverse transcription-polymerase chain reaction assay validated in vitro and with clinical specimens. J Clin Microbiol. 2020 Mar 4.

### Sensitive and specific detection of COVID-19

An experimental model containing synthetic sequences targeted by the SARS-CoV-2 detection kit was serially diluted and seven dilution points were assessed in triplicate. A total of 1ng of human RNA was added to each replicate. The results indicated a robust and specific detection of SARS-CoV-2 sequences down to 0.8 copies per  $\mu$ l of positive control (5 copies per 25 $\mu$ l reaction) of the ORF1ab gene and down to 0.9 copies per  $\mu$ l of positive control (7 copies per 25 $\mu$ l reaction) of the Nucleocapside (N) gene in all tested samples. Further dilutions showed an extremely sensitive but stochastic detection down to 0.25 copies per  $\mu$ l of positive control (2 copies per 25 $\mu$ l reaction) for both genes (Figure 3). In parallel, a total of 15 controls containing only human RNA were tested as negative controls and no false positives were observed.

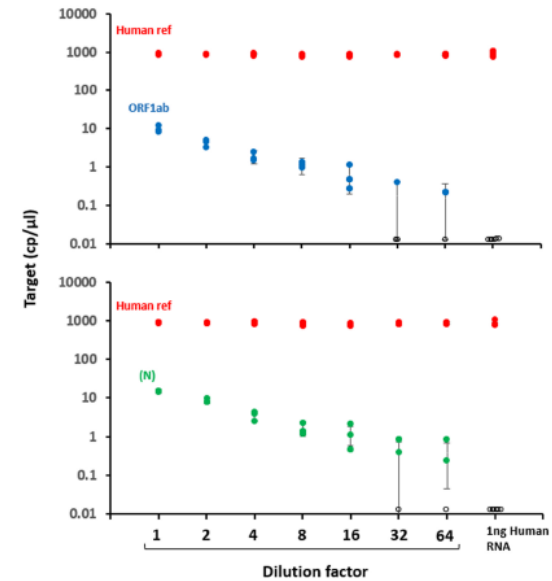
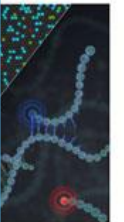


Figure 3: Sensitivity of the 3-color RUO RT Crystal Digital PCR™ kit for SARS-CoV-2 detection. Serial dilutions of SARS-CoV-2 synthetic targets were assayed in triplicate in a background of 1ng of human RNA. A total of 8 $\mu$ l of positive controls was added to each 25 $\mu$ l reaction. The vertical bars represent the theoretical 95% Poisson confidence intervals for the pool of 3 replicates. The empty circles represent replicates where SARS-CoV-2 sequences were not detected.

LA  
 Quantify Drop-Off  
 PCR assays  
 Crystal Miner™





# LEARNING CENTER: [www.gene-pi.com](http://www.gene-pi.com)

GENE- $\pi$

HOME TUTORIALS HOW TO STATISTICAL TOOLS TRAINING COMMUNITY

## DIGITAL PCR

Learn, train and experiment with cutting-edge tools and methods

What do you want to learn ?...

## LAUNCH IN MARCH 2019:



### 3 tutorials

- Rare Mutation Detection
- CNV
- Drop-off Assay



### 1 video



### 14 how to's



### 3 memos



### 3 online statistical tools

- Poisson Law
- CNV
- Limit of Blank/Limit of Detection





## **SPECIAL THANK YOU TO:**

Pr. Michael Drancourt

Dr. Amar Bouam

Romain Parillaud

## **THANK YOU FOR YOUR ATTENTION!**

### **ANY QUESTIONS?**

Kimberley.Gutierrez@stilla.fr

For more information on product and workflow, visit our website at

**[www.stillatechnologies.com](http://www.stillatechnologies.com)**



